



HAL
open science

Gut microbiota imbalance and colorectal cancer.

Johan Gagnière, Jennifer Raisch, Julie Veziat, Nicolas Barnich, Richard Bonnet, Emmanuel Buc, Marie-Agnès Bringer, Denis Pezet, Mathilde Bonnet

► **To cite this version:**

Johan Gagnière, Jennifer Raisch, Julie Veziat, Nicolas Barnich, Richard Bonnet, et al.. Gut microbiota imbalance and colorectal cancer.. World Journal of Gastroenterology, 2016, 22 (2), pp.501-18. 10.3748/wjg.v22.i2.501 . pasteur-01351170

HAL Id: pasteur-01351170

<https://riip.hal.science/pasteur-01351170v1>

Submitted on 2 Aug 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License



2016 Colorectal Cancer: Global view

Gut microbiota imbalance and colorectal cancer

Johan Gagnière, Jennifer Raisch, Julie Veziat, Nicolas Barnich, Richard Bonnet, Emmanuel Buc, Marie-Agnès Bringer, Denis Pezet, Mathilde Bonnet

Johan Gagnière, Jennifer Raisch, Julie Veziat, Nicolas Barnich, Richard Bonnet, Emmanuel Buc, Marie-Agnès Bringer, Denis Pezet, Mathilde Bonnet, Clermont Université, UMR 1071 Inserm/Université d'Auvergne, 63000 Clermont-Ferrand, France

Johan Gagnière, Jennifer Raisch, Julie Veziat, Nicolas Barnich, Richard Bonnet, Emmanuel Buc, Marie-Agnès Bringer, Denis Pezet, Mathilde Bonnet, INRA, USC-2018, 63000 Clermont-Ferrand, France

Johan Gagnière, Julie Veziat, Emmanuel Buc, Denis Pezet, Chirurgie Digestive, Centre Hospitalier Universitaire, 63000 Clermont-Ferrand, France

Richard Bonnet, Bactériologie, Centre Hospitalier Universitaire, 63000 Clermont-Ferrand, France

Jennifer Raisch, Laboratoire d'Immunologie, Institut Armand Frappier, Laval H7V 1B7, Canada

Marie-Agnès Bringer, INRA UMR 1324, CNRS UMR 6265, Université de Bourgogne, Centre des Sciences du Goût et de l'Alimentation, Eye Nutrition and Signalling Research Group, 21000 Dijon, France

Author contributions: Gagnière J, Raisch J and Veziat J organized and wrote the manuscript; Barnich N, Bonnet R, Buc E, Bringer MA and Pezet D supervised the writing of the manuscript; Bonnet M organized, wrote and supervised the writing and the manuscript.

Supported by Inserm and Université d'Auvergne (UMR 1071), INRA (USC-2018); and grants from "Conseil régional d'Auvergne", "Nuovo Soldati Foundation for Cancer Research" and "Fondation pour la recherche médicale".

Conflict-of-interest statement: None of the authors have any conflicts of interest to declare.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this

work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Mathilde Bonnet, PhD, M2iSH "Microbes, intestin, inflammation et Susceptibilité de l'Hôte" UMR 1071 Inserm/Université d'Auvergne USC INRA 2018, Centre Biomédical de Recherche et Valorisation, 28 Place Henri Dunant, 63000 Clermont-Ferrand, France. mathilde.bonnet@udamail.fr
Telephone: +33-4-7318381
Fax: +33-4-73178371

Received: April 28, 2015

Peer-review started: May 6, 2015

First decision: August 25, 2015

Revised: September 6, 2015

Accepted: October 17, 2015

Article in press: October 20, 2015

Published online: January 14, 2016

Abstract

The gut microbiota acts as a real organ. The symbiotic interactions between resident micro-organisms and the digestive tract highly contribute to maintain the gut homeostasis. However, alterations to the microbiome caused by environmental changes (*e.g.*, infection, diet and/or lifestyle) can disturb this symbiotic relationship and promote disease, such as inflammatory bowel diseases and cancer. Colorectal cancer is a complex association of tumoral cells, non-neoplastic cells and a large amount of micro-organisms, and the involvement of the microbiota in colorectal carcinogenesis is becoming increasingly clear. Indeed, many changes in the bacterial composition of the gut microbiota have been reported in colorectal cancer, suggesting a major role of dysbiosis in colorectal carcinogenesis. Some bacterial species have been identified and suspected to play a role in colorectal carcinogenesis, such as *Streptococcus bovis*, *Helicobacter pylori*, *Bacteroides*

fragilis, *Enterococcus faecalis*, *Clostridium septicum*, *Fusobacterium* spp. and *Escherichia coli*. The potential pro-carcinogenic effects of these bacteria are now better understood. In this review, we discuss the possible links between the bacterial microbiota and colorectal carcinogenesis, focusing on dysbiosis and the potential pro-carcinogenic properties of bacteria, such as genotoxicity and other virulence factors, inflammation, host defenses modulation, bacterial-derived metabolism, oxidative stress and anti-oxidative defenses modulation. We lastly describe how bacterial microbiota modifications could represent novel prognosis markers and/or targets for innovative therapeutic strategies.

Key words: Colorectal cancer; Gut microbiota; Dysbiosis; Cyclomodulin; Oxidative stress

© **The Author(s) 2016.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The gut microbiota acts as a real organ and many changes in its composition have been reported in colorectal cancer. The pro-carcinogenic properties of bacteria are now better understood. In this review, we discuss possible links between the bacterial microbiota and colorectal carcinogenesis, focusing on the dysbiosis-causing and pro-carcinogenic properties of bacteria, such as genotoxicity, inflammation, and oxidative stress. We lastly detail how microbiota modifications may represent novel prognosis markers and/or targets for innovative therapeutic strategies.

Gagnière J, Raisch J, Veziat J, Barnich N, Bonnet R, Buc E, Bringer MA, Pezet D, Bonnet M. Gut microbiota imbalance and colorectal cancer. *World J Gastroenterol* 2016; 22(2): 501-518 Available from: URL: <http://www.wjgnet.com/1007-9327/full/v22/i2/501.htm> DOI: <http://dx.doi.org/10.3748/wjg.v22.i2.501>

INTRODUCTION

Colorectal cancer (CRC) is a complex association of tumoral cells, non-neoplastic cells (*i.e.*, stromal cells) and a large number of micro-organisms. The microbiota may be linked to carcinogenesis *via* various mechanisms and could lead to the development of novel prognosis markers and/or targets for innovative therapeutic strategies.

GUT MICROBIOTA IN HEALTH

What is the gut microbiota?

Approximately 100 trillion micro-organisms (including bacteria, viruses and fungi) reside in the adult human gut and constitute the microbiota^[1,2]. The composition of the microbiota is rather stable along the length of the gut, but the absolute number of micro-organisms varies considerably between the mouth and rectum^[3].

The gut microbiota consistently differs among between individuals. It is acquired during the first stages of life *via* the commensal flora from the mother's skin, vagina and feces, and matures primarily during the first two years. Microbiota development is the result of interactions between physiological process in the host and micro-organisms that are introduced from the environment^[4-6]. After the initial stages, the microbiota stabilizes and maintains a consistent composition, despite some fluctuations throughout adulthood in response to environmental, developmental and pathological events^[7,8]. In the elderly, the microbiota composition changes gradually but can maintain similar physiological functions^[9-12]. The early acquisition of a diverse and balanced microbiota is likely critical for the development and maturation of a healthy immune system, as suggested by immune abnormalities in germ-free animals raised in bacteria-free conditions^[4,13]. The colon is colonized by approximately 10³ different microbial species and this colonic microbiota is mostly represented by bacteria^[7,9,14]. Indeed, the colon contains approximately 10¹⁴ bacteria (70% of the host's microorganisms)^[15,16]. This review will therefore focus on the impact of bacteria in CRC.

Most bacteria cannot be cultured, but modern molecular approaches can be used to identify and classify bacteria such as 16S ribosomal RNA (16S rRNA) sequencing from feces or digestive tissues can be used to identify and classify bacteria. The microbiota can be divided according to location in the gut. Specifically, microbes in the lumen are referred to "luminal flora", whereas microbes that penetrate the mucosal layer overlying the intestinal epithelium are referred to "mucosa-associated flora"^[16]. Indeed, thick mucus layers protect enterocytes from excessive exposure to microorganisms and dietary antigens along the length of the intestine, particularly in the colon, thus preventing hypersensitivity responses^[17]. Moreover, the ratio of anaerobes to aerobes is lower at the mucosal surfaces than in the lumen. In addition, the collection of "fecal flora" from the feces is a non-invasive technique that facilitates sampling of colonic microbiota. These bacteria are representative of distal colonic colonization but differ from proximal "associated" flora^[18]. It is essential to note that the composition of murine gut microbiota is quite similar to that in humans, lending translational relevance to mouse experimental models of gastrointestinal disease^[19,20]. Actually, more than 50 different phyla and 500 bacterial species may comprise the human normal commensal gut microbiota. Although the exact number of species and the amount of variability among individuals remain to be characterized^[21,22], these factors are likely highly dependent on lifestyle, diet and host genotype^[23,24]. Some bacterial species are regularly recovered from different individuals, and the human gut microbiota is dominated by 3 primary phyla: *Firmicutes* (30%-50%), *Bacteroidetes* (20%-40%)

and *Actinobacteria* (1%-10%). Strict anaerobes, including *Bacteroides*, *Eubacterium*, *Bifidobacterium*, *Fusobacterium*, *Peptostreptococcus* and *Atopobium*^[25], represent a major portion of the gut microbiota, whereas facultative anaerobes, such as *Lactobacilli*, *Enterococci*, *Streptococci* and *Enterobacteriaceae*, constitute a minority (about 1000-fold lower levels). The fact that composition significantly vary along the gut should also be highlighted, especially given that *Bacteroidetes* and *Actinobacteria* represent more than 90% of bacterial phyla in the colon but only 50% in the small intestine, which contains approximately 40% *Firmicutes* species^[26].

Microbiota and gut homeostasis

The gut microbiota constitutes a natural defensive barrier to infection. Moreover, the microbiota is involved in numerous protective, structural and metabolic roles in the intestinal epithelium and plays a large role in maintaining gut homeostasis. The microbiota is involved in several physiological functions^[27]. The impact of enteric bacteria on intestinal physiology has been studied primarily in germ-free animals raised in bacteria-free conditions. Such animals are more susceptible to infections and have reduced vascularity, digestive enzyme activity, muscle wall thickness, cytokine production and serum immunoglobulin levels, smaller Peyer's patches and fewer intraepithelial lymphocytes^[28]. The reconstitution of a gut microbiota in germ-free mice is sufficient to restore the mucosal immune system^[29] and affects the expression of various host genes that can impact nutrient uptake, metabolism, angiogenesis, mucosal barrier function and development of the intestinal nervous system^[30]. Moreover, commensal bacteria influence the normal development and function of the mucosal immune system^[31,32], such as the humoral components^[33]. These bacteria also modify T-cell repertoires and T-helper-cell cytokine profiles^[34,35]. Such data support a possible influence of gut microbiota composition on individual variations in immunity.

The structural role of gut microbiota on the intestinal epithelium is increasingly evident and has been studied primarily in germ-free mice. These animals present longer intestinal villi, associated with crypt atrophy, slower renewal of epithelial cells and decrease of angiogenesis phenomenon^[3]. Furthermore, it has been reported that mucosa and muscle wall thickness were decreased in these mice^[28] and that microbiota enhanced crypt cell turnover in a CRC-predisposed mouse model^[36].

Furthermore, the gut microbiota is involved in metabolic functions^[27]. For example, the microbiota can participate in (1) anaerobic carbohydrate fermentation through the production of CO₂, H₂, CH₄ and short-chain fatty acids (*e.g.*, butyrate, propionate and acetate); and (2) proteolytic fermentation *via* metabolites such as phenolic compounds, amines,

ammonia, N-nitroso compounds and indoles. These effects can impact on gene expression, intestinal epithelial cell differentiation and proliferation, and can also mediate vitamin synthesis, ion absorption and mucus production^[27,35,37,38]. This complex metabolic activity also increases the yield and storage of energy from dietary sources, regulates fat storage, helps to provide absorbable substrates for both the host and the microbiota, and is involved in bacterial growth and proliferation^[37-40]. Some of produced metabolites, especially during proteolytic fermentation, can be toxic to the host^[27,41,42].

In addition to immune, structural, and metabolic functions, the commensal microbiota inhibits gut colonization of intruding pathogens and ensures "colonization resistance" or "microbial interference"^[43]. The involved mechanisms of these effects remain unclear but likely involve competition with adhesion receptors, stabilization of the gut mucosal barrier, competition for nutrients and the production of anti-microbial substances^[27]. Indeed, alterations in colonization resistance due to, *e.g.*, pathogens or antibiotics treatment, probably increase the risk of gastrointestinal affections.

MICROBIOTA AND COLORECTAL CANCER

Geographic variability of the incidence of CRC highly suggests the involvement of certain environmental risk factors, such as high-fat diets, obesity or living in a Western country^[44,45]. Moreover, Knudson's two-hit hypothesis suggests that host factors play an important role in the predisposition to carcinogenesis. In this scenario, a second environmental hit can lead to uncontrolled cellular proliferation^[46]. Indeed, growing attention has been given to the role of microbial infection in carcinogenesis in recent decades, and microbes are suspected to be involved in approximately 20% of cancers^[47], especially CRC^[48]. Concerning digestive cancers, even if some pathogens, such as *Helicobacter pylori* (*H. pylori*), have been directly and strongly linked to gastric cancer^[49], possible infectious cause in CRC remains controversial. It is becoming increasingly clear, however, that pathogens play a role in colorectal carcinogenesis^[50]. Interestingly, bacteria levels in the colon are one million-fold higher than in the small intestine, and approximately 12-fold more cancers develop in the former than in the latter, suggesting a potential role of gut microbiota in colorectal carcinogenesis^[51]. The first observation linking gut microbiota with CRC was reported in 1975 in germ-free rats that developed less chemically induced colorectal tumor than conventional rats^[52]. These results have been reproduced in CRC-predisposed mice^[36,53]. Contrary to gastric carcinogenesis, which seems to result from a single pathogen, the following differing hypotheses have emerged to explain the

contribution of bacteria to CRC: (1) a dysbiotic microbial community with pro-carcinogenic features are capable of remodeling the microbiome as a whole to drive pro-inflammatory responses and epithelial cell transformation, leading to cancer; and (2) the “driver-passenger” theory, wherein intestinal bacteria, termed “bacteria drivers”, initiate CRC by inducing epithelial DNA damage and tumorigenesis, in turn promoting the proliferation of passenger bacteria that have a growth advantage in the tumoral microenvironment^[50,54]. Studies in mouse models of altered immune and inflammatory responses suggest that dysbiosis could be sufficient to promote cancer^[55,56]. It is thus likely that the immune system is a key factor in the interactions between the gut microbiota and CRC. In addition to the impact of specific pathogens on carcinogenesis, the high redundancy of gut microbiota at a metagenomic level suggests that dysbiosis could exert pro-carcinogenic effects^[57]. These properties could be due to interactions between different emergent bacterial strains activating similar pathways. However, the mechanisms that contribute to dysbiosis and to alterations in microbial richness are not well understood, and it is unknown whether dysbiosis is a cause or a consequence of CRC. Indeed, the CRC microenvironment is characterized by host-derived immune and inflammatory responses that could impact on microbial regulation, alter microbiota composition, and favor the outgrowth of specific bacteria that potentially have carcinogenic effects^[58]. Dysbiosis in CRC could thus result in selection of microbiota composition *via* a tumor-linked microenvironment, with the emergence of “keystone pathogens” that have strong effects on bacterial composition and subsequently amplify dysbiosis^[59].

With the 16S rRNA sequencing of bacteria from the feces or digestive tissues, numerous studies have reported colonic dysbiosis in patients presenting with CRC^[18,60-64]. At present, there is no consensus with respect to the modifications observed in CRC, which are listed in Table 1. However, some bacterial species have been identified and are suspected to play a role in colorectal carcinogenesis^[27,54]. These species primarily include *Streptococcus bovis* (*S. bovis*)^[62,65,66], *H. pylori*^[67-69], *Bacteroides fragilis* (*B. fragilis*)^[61,62,70-72], *Enterococcus faecalis* (*E. faecalis*)^[62,73], *Clostridium septicum* (*C. septicum*)^[74-76], *Fusobacterium* spp.^[77-79] and *Escherichia coli* (*E. coli*)^[80-82].

S. bovis/gallolyticus

S. bovis was the first bacteria indirectly associated with CRC. Indeed, McCoy *et al*^[83] reported the first recognized case of enterococcal endocarditis associated with CRC in 1951. This association was later confirmed by Hoppes *et al*^[84], who reported that approximately two-thirds of *S. bovis* endocarditis cases were associated with gastrointestinal disease. In addition, Klein *et al*^[85] reported a 5-fold increased incidence of CRC in patients presenting such endocarditis.

Interestingly, in this study, most of patients had asymptomatic colorectal tumors that were occasionally benign adenomas, suggesting the involvement of *S. bovis* at an early step of CRC development. However, there are contrasting results in studies of the association between *S. bovis* and CRC; such differences could be due to variations in the collection of feces, sample processing, bacterial culturing or the molecular analysis of fecal samples^[66,86,87].

Studies that confirmed the association between *S. bovis* infection and CRC reported prevalences from 33% to 100% of *S. bovis* with underlying CRC^[88]. It was shown that the mucosal detection of *S. bovis* could be a better tool than fecal level to assess its presence in patients with CRC^[27,89]. More recently, Abdulmir *et al*^[90] used molecular techniques to show increased *S. bovis* in colorectal adenomas and CRC tissues, strengthening the possible involvement of this pathogen in colorectal carcinogenesis. The mechanisms behind this link remain unclear, but these three hypotheses have been reported: (1) *S. bovis* adheres to both normal epithelial and neoplastic cells; (2) this species attains a competitive growth advantage in a tumor microenvironment by foraging on tumor metabolites; and (3) *S. bovis* induces inflammation and/or pro-carcinogenic pathways, leading to tumor progression, especially from pre-malignant tumors^[27]. Thus, the fact that increase in the numbers of *S. bovis* probably occurs at an early step in colorectal carcinogenesis could lead its use for early detection of CRC^[91]. Moreover, these results directly impact clinical practice, leading to recommended colorectal endoscopic exploration in all patients presenting with *S. bovis* infection.

H. pylori

H. pylori has been directly and strongly linked to gastric cancer^[49] and has recently been classified as a carcinogen of the gastrointestinal tract by the International Agency for Research on Cancer. The role of *H. pylori* in colorectal carcinogenesis remains controversial due to differences between studies with respect to the *H. pylori* strains and their specific virulence factors. A meta-analysis of 11 studies conducted between 1991 and 2002 was published by Zumkeller *et al*^[69] and reported a 1.4-fold increased risk of CRC in patients presenting with a *H. pylori* infection. More recently, Guo *et al*^[92] reported a statistical association between *H. pylori* and colorectal adenomas in a meta-analysis of 7679 Asian patients. This result suggested a carcinogenic role of *H. pylori* at an early stage of carcinogenesis. The presence of high levels of *H. pylori* has been reported in CRC tissue using a specific 16S rDNA polymerase chain reaction (PCR) assay and pyrosequencing^[67]. Jones *et al*^[68] analyzed paraffin-embedded tissue blocks of normal colonic mucosa, adenomas and adenocarcinomas from 180 patients. The results indicated a significant

Table 1 Summary of 16S rRNA sequencing and qPCR analyses of colonic microbiota variations in colorectal cancer

Variation in CRC	Phyla	Genus/species	Population	Ref.
Fecal flora				
↑		<i>Enterococcus Faecalis</i>	20 CRC/17 C	[73]
↑	Proteobacteria	<i>Porphyromonas/Escherichia/Shigella Enterococcus/ Streptococcus/Peptostreptococcus</i>	46 CRC/56 C	[62]
		<i>Bacteroides fragilis</i>		
↑		<i>Bacteroides/Prevotella</i>	60 CRC/119 C	[61]
↑		<i>Peptostreptococcus/Mogibacterium</i>	21 CRC/22 C	[18]
		<i>Anaerococcus/Slakia/Paraprevotella</i>		
		<i>Anaerotruncus/Collinsella/Desulfovibrio</i>		
		<i>Eubacterium/Porphyromonas</i>		
↑	Bacteroidetes	<i>Atopobium/Porphyromonas</i>	47 CRC/94 C	[63]
	Fusobacteria	<i>Fusobacterium</i>		
↑	Fusobacteria	<i>Fusobacterium/Bacteroides</i>	19 CRC/20 C	[64]
↑		<i>Bacteroides/Fusobacterium</i>	46 CRC/63 C	[214]
		<i>Alistipes/Escherichia/Parvoimonas/Bilophila</i>		
		<i>Faecalibacterium prauznitsii</i>	20 CRC/17 C	[73]
↓	Bacteroidetes	<i>Bacteroides vulgatus/Bacteroides uniformis</i>	46 CRC/56 C	[62]
		<i>Roseburia/Butyrate-producing bacteria</i>		
↓		<i>Faecalibacterium prauznitsii/Roseburia</i>	19 CRC/20 C	[64]
↓	Firmicutes (clostridia)	<i>Ruminococcus</i>	47 CRC/94 C	[63]
↓		<i>Ruminococcus/Bifidobacterium/Streptococcus</i>	46 CRC/63 C	[214]
Tumor-associated flora				
↑	Bacteroidetes	<i>Coriobacteriae/Roseburia</i>	6 CRC/6 AM	[14]
		<i>Fusobacterium/Faecalibacterium</i>		
		<i>Butyrate-producing bacteria</i>		
↑		<i>Bacteroides</i>	22 CRC/22 C	[61]
↑		<i>Fusobacterium</i>	95 CRC/95 AM	[106]
↑	Bacteroidetes	<i>Bacteroides/Prevotella/Streptococcus</i>	27 CRC	[18]
	Fusobacteria	<i>Fusobacterium/Peptostreptococcus</i>	27 intestinal lumen	
	Proteobacteria	<i>Morganella/Porphyromonas</i>		
↑		<i>Fusobacterium</i>	55 CRC/55 AM	[109]
↓	Firmicutes	<i>Shigella/Citrobacter/Serratia/Salmonella</i>	6 CRC/6 AM	[14]
↓	Firmicutes	<i>Lactobacillus/Roseburia/Pseudobutyrvibrio</i>	27 CRC	[18]
			27 intestinal lumen	
↓	Bacteroidetes		95 CRC/95 AM	[106]
	Firmicutes (clostridia)			
Mucosa-associated flora				
↑		<i>Fusobacterium/Porphyromonas</i>	32 CRC	[18]
		<i>Peptostreptococcaceae/Gemella</i>	22 C (swab)	
		<i>Mogibacterium/Klebsiella</i>		
↓		<i>Faecalibacterium/Blautia/Anaerostipes</i>	32 CRC	[18]
		<i>Lachospira/Bifidobacterium</i>	22 C (swab)	

CRC: Colorectal cancer; C: Control patients; AM: Adjacent tumor, normal mucosa of patients with CRC.

increase in colonic mucosa-associated colonization by *H. pylori* in adenomas and adenocarcinomas compared to normal mucosa^[68].

Although the role of *H. pylori* in gastric carcinogenesis has been better studied and described than its involvement in colorectal carcinogenesis, some hypotheses can be extrapolated from the pathophysiology of bacteria-linked gastric cancer. Bacterial cytotoxin-associated gene A (CagA) and vacuolating cytotoxin A (VacA) are encoded by pathogenicity islands in some *H. pylori* strains and may induce the activation of inflammation pathways and cellular proliferation in gastric cancer^[93]. Shmueli *et al.*^[94] reported that patients presenting with seropositivity for CagA-positive *H. pylori* strains had a significantly increased risk of CRC. Another hypothesis is the direct and indirect production of pro-oxidative reactive oxygen

and nitrogen species by some *H. pylori* strains that could participate to colorectal carcinogenesis^[49].

B. fragilis

B. fragilis are common anaerobic bacteria that are detected in up to 80% of children and adults but which represent less than 1% of gut microbiota^[95,96]. There are two molecular subtypes of *B. fragilis*, nontoxigenic or enterotoxigenic, with the latter known to be responsible in diarrheal illnesses in humans, especially children^[97]. Enterotoxigenic *B. fragilis* is easily detected in fecal samples of up to 40% of healthy adults, facilitating studies on the role of these bacteria in gastrointestinal diseases^[98]. Indeed, some studies have reported increased colonic colonization by *B. fragilis* in patients presenting with CRC^[61,62]. For example, Sobhani *et al.*^[61] performed pyrosequencing

on stools from 179 patients with CRC or controls who had a normal colonoscopy. The results indicated more colonization by *B. fragilis* in patients presenting with CRC than in control patients^[61]. Most of enterotoxigenic *B. fragilis* strains detected in mucosal samples from patients with CRC harbored the *bft* gene, which encodes the bacterial toxin *B. fragilis* toxin (BFT). This toxin is involved in the pathogenicity of enterotoxigenic *B. fragilis*^[99,100]. Indeed, the involvement of BFT in colorectal carcinogenesis is becoming increasingly evident as this toxin directly affects pathways that lead to increased cell proliferation, the epithelial release of pro-inflammatory effectors, and DNA damage in *in vitro* and CRC-predisposed mouse models^[70-72].

E. faecalis

E. faecalis is a facultative anaerobic commensal bacterium of the oral cavity and the gastrointestinal tract. Recently, this species has emerged as a human pathogen^[101]. Balamurugan *et al.*^[73] performed 16S rRNA real-time PCR from the feces of patients with CRC and healthy volunteers. The authors reported significantly higher *E. faecalis* fecal populations in patients with CRC compared to healthy controls^[62,73]. These results were confirmed more recently by Wang *et al.*^[62]. The mechanisms linking *E. faecalis* to colorectal carcinogenesis remain unclear, but the production of pro-oxidative reactive oxygen species (ROS) by *E. faecalis* has been described in cellular and animal models^[102,103]. Moreover, *E. faecalis* can trigger colitis, dysplasia and CRC in a susceptible interleukin (IL)-10^{-/-} mouse model^[104].

C. septicum

C. septicum is a rare cause of bacteremia (less than 1%). As for *S. bovis* endocarditis, *C. septicum* infections have been clinically linked to CRC^[76]. Indeed, it has been reported that gastrointestinal disease and/or colorectal malignancies can be found in up to 40% of patients presenting with *C. septicum* infections^[74,75]. Hermsen *et al.*^[75] reported an analysis of 320 cases of *C. septicum* infections, of which more than 40% had a gastrointestinal origin, primarily malignant. The underlying mechanisms of this association remain unknown, but one hypothesis is that the hypoxic and acidic tumor microenvironment favors the germination of *C. septicum* spores *via* ingestion of contaminated food^[27,105]. However, no direct involvement of *C. septicum* in colorectal carcinogenesis has been well defined.

Fusobacterium nucleatum

Fusobacterium nucleatum (*F. nucleatum*) is an anaerobic Gram-negative pathogenic bacterium that recently emerged as a potential candidate for CRC susceptibility. Indeed, recent metagenomic analyses using whole-genome and bacterial 16S rRNA sequencing revealed an enrichment of *F. nucleatum*

in colonic tumor-associated microbiota compared to adjacent normal mucosa in patients with CRC^[106,107]. Moreover, other studies found that luminal *F. nucleatum* colonization was higher in patients presenting with colorectal adenomas compared to healthy patients. An increase of *F. nucleatum* colonization in adenomas compared to adjacent colonic normal mucosa has also been reported^[77,78]. These data strongly suggest that, more than simply being associated with CRC, *F. nucleatum* likely acts at the early steps of colorectal carcinogenesis promotion. This effect may be mediated *via* its FadA adhesion and activates the Wnt/ β -catenin pathway^[79]. This hypothesis has been verified in a CRC-predisposed adenomatous polyposis coli (APC)^{min/+} mouse model. In this animal model, infection with *F. nucleatum* increased tumor multiplicity and selectively recruited tumor-infiltrating myeloid cells, which can promote tumor progression^[77]. Interestingly, *F. nucleatum* is detected in up to 80% of tumor samples. The increased colonic colonization of this species has been correlated with CRC stage, as has been described for other pathogens^[108,109]. Indeed, Viljoen *et al.*^[109] identified significant associations between high-level colonization by *F. nucleatum* and advanced stage III-IV CRC, as well as with microsatellite instability tumor phenotype. These results could lead to further studies that explore (1) the role of *F. nucleatum* in modulating the DNA repair systems involved in colorectal carcinogenesis; and (2) the potential pathogenic interactions between different pathogenic bacterial species.

E. coli

Although *E. coli* is a commensal bacteria of the human microbiota and represents the most common cultivable, Gram-negative, aero-anaerobic bacteria in the gut, various studies have demonstrated a clear link between mucosa-adherent *E. coli* and CRC^[80-82]. *E. coli* belonging to phylogroups B2 and D comprise most of the pathogenic strains that express virulence factors, and some of these species are involved in chronic inflammatory bowel diseases, which are known risk factors for CRC^[110,111].

Some studies have reported higher levels of colonic colonization by mucosa-associated *E. coli* in patients with CRC compared to healthy patients^[80,81]. For example, Swidsinski *et al.*^[81] performed 16S PCR using colonic mucosa tissue samples and detected *E. coli* in only 3% of healthy patient biopsies. However, *E. coli* was detected and predominant in 62% and 77% of patients presenting with adenomas and carcinomas, respectively^[81]. A few years later, Martin *et al.*^[80] reported that more than 70% of mucosa samples from patients with CRC were colonized by bacteria, generally *E. coli*. Increased mucosa- and tumor-associated *E. coli* colonization in CRC patients has been confirmed by several studies, strongly implicating *E. coli* in colorectal carcinogenesis^[82,108,112,113].

CRC samples commonly exhibit colonic mucosal

E. coli that could express genes (*afa*, *lpfA*, *eae* and/or cyclomodulin toxins) that confer characteristics that are relevant to pathogenesis, including M-cell translocation, angiogenesis and genotoxicity^[112,114-117]. Cyclomodulins such as colibactin, are toxins that interfere with the eukaryotic cell cycle and induce DNA damage and genomic instability, which are involved in colorectal carcinogenesis^[118,119]. A pro-carcinogenic effect of B2 phylogroup colibactin-producing *E. coli* has been confirmed in mouse models. Specifically, these bacteria are involved in inflammation pathways and cell proliferation fueling^[82,108,120]. Moreover, possible interactions between *E. coli* and DNA repair system involved in colorectal carcinogenesis, such as the DNA mismatch repair (MMR) system, have been described^[112,121]. Interestingly, these B2 colibactin-producing *E. coli* are preferentially detected in patients with CRC^[82,108,115,120] and were more prevalent in the mucosa of patients presenting with advanced stage III/IV CRC than in those with stage I CRC^[108]. These results suggest that *E. coli* could serve as a prognostic factor in CRC.

SUSPECTED INVOLVED MECHANISMS

As described above, some bacterial species may be involved in colorectal carcinogenesis. Many suspected involved mechanisms, occasionally shared by different species, have been described. These mechanisms include bacterial-derived genotoxins, microbial-derived metabolism, the modulation of host defenses and inflammation pathways, oxidative stress induction, and anti-oxidative defense regulation (Figure 1).

Bacterial-derived genotoxins and other bacterial virulence factors

During their phylogenetic evolution, bacteria progressively acquired virulence factors that conferred pathogenicity. For example, bacteria developed the ability to penetrate the gut mucosal barrier, as well as to adhere to and invade intestinal epithelial cells, specifically through the use of flagellum, pili, and adhesins^[110,122,123]. Most of the disease-promoting and pro-carcinogenic effects of pathogens depend on these virulence factors^[58]. Pathogens can interact with adhesion molecules. For example, *F. nucleatum* uses the FadA virulence factor to adhere to and invade cells^[124], thereby activating β -catenin signaling pathway and promoting CRC^[79]. In the same way, certain CRC-associated *E. coli* strains have acquired virulence factors, such as the *afa* and *eae* adhesins, which confer the ability to adhere to and invade the intestinal epithelium^[112,116].

Toxins may be involved in colorectal carcinogenesis by modulating certain host-derived signaling pathways, resulting in the activation of carcinogenesis-promoting pathways. For example, CagA or VacA are produced by some *H. pylori* strains and increase

inflammation and cancer rates^[125,126]. *B. fragilis* has been associated with CRC^[71,127], and BFT is another well-described example of a bacteria-derived toxin involved in colorectal carcinogenesis. BFT is a 20 kDa zinc-dependent metalloprotease toxin that is grouped into three isotypes, namely, BFT-1, BFT-2 and BFT-3^[100]. These toxins are encoded by a unique *B. fragilis*-specific gene, which was described by Moncrief *et al.*^[128] in 1995. A recent study reported that mucosa-associated BFT-producing *B. fragilis* were more prevalent in late-stage CRC, suggesting possible role of BFT in CRC promotion and progression^[99]. At the molecular level, BFT binds to a specific colonic epithelial receptor, activating the Wnt and nuclear factor-kappa B (NF- κ B) pathways. These effects lead to increased cell proliferation, the epithelial release of pro-inflammatory mediators, and the induction of DNA damage^[97,129-132]. Furthermore, in the CRC-predisposed multiple intestinal neoplasia APC^{min/+} mouse model, BFT promotes IL-17-dependent carcinogenesis^[127].

Other toxins, named cyclomodulins [cytotoxic distending toxin (CDT), colibactin, cytotoxic necrotizing factor and cycle inhibiting factor], can induce DNA damage, interfere with the cell cycle and/or apoptosis^[82,118,127,133-136]. Only CDT and colibactin can directly damage DNA and are linked genomic instability; these proteins are therefore considered true genotoxins^[118,133]. Both CDT and colibactin induce double-strand DNA breaks, activate the ataxia telangiectasia mutated (ATM)-checkpoint kinase 2 signaling pathway, and lead to H2AX histone phosphorylation. These effects result in transient G2/M cell cycle arrest and cell swelling.

Most of the Gram-negative bacteria that are involved in CRC produce CDT, which is by far the most well-characterized bacterial-derived genotoxin^[137]. The CdtA and CdtC subunits permit the interaction between pathogens and host cells; subsequently, the cytoplasmic CdtB subunit can translocate to the nucleus, act as a DNase and damage DNA^[133]. CDT also favors persistent gut colonization and induces the production of pro-inflammatory molecules, such as NF- κ B, tumor necrosis factor (TNF)- α , IL-6 and cyclooxygenase (COX) 2. These factors are involved in many carcinogenic processes. CDT has also been reported to induce dysplasia in a *Helicobacter hepaticus*-linked hepatocarcinoma mouse model^[138,139]. When combined, DNA damage, interference with the cell cycle and the modulation of pro-inflammatory pathways can lead to mutations that are involved in genomic instability in CRC.

Colibactin is another bacterial-derived genotoxin that has recently attracted attention. It was first described in 2006 by Nougayrède *et al.*^[119] but has not yet been isolated and purified. Colibactin synthesis and activation requires complex machinery that involves three non-ribosomal peptide megasynthases (NRPS), three polyketide megasynthases (PKS), two NRPS/PKS hybrids and at least eight of nine accessory enzymes encoded in the *pks* island^[119]. The *pks* island is found

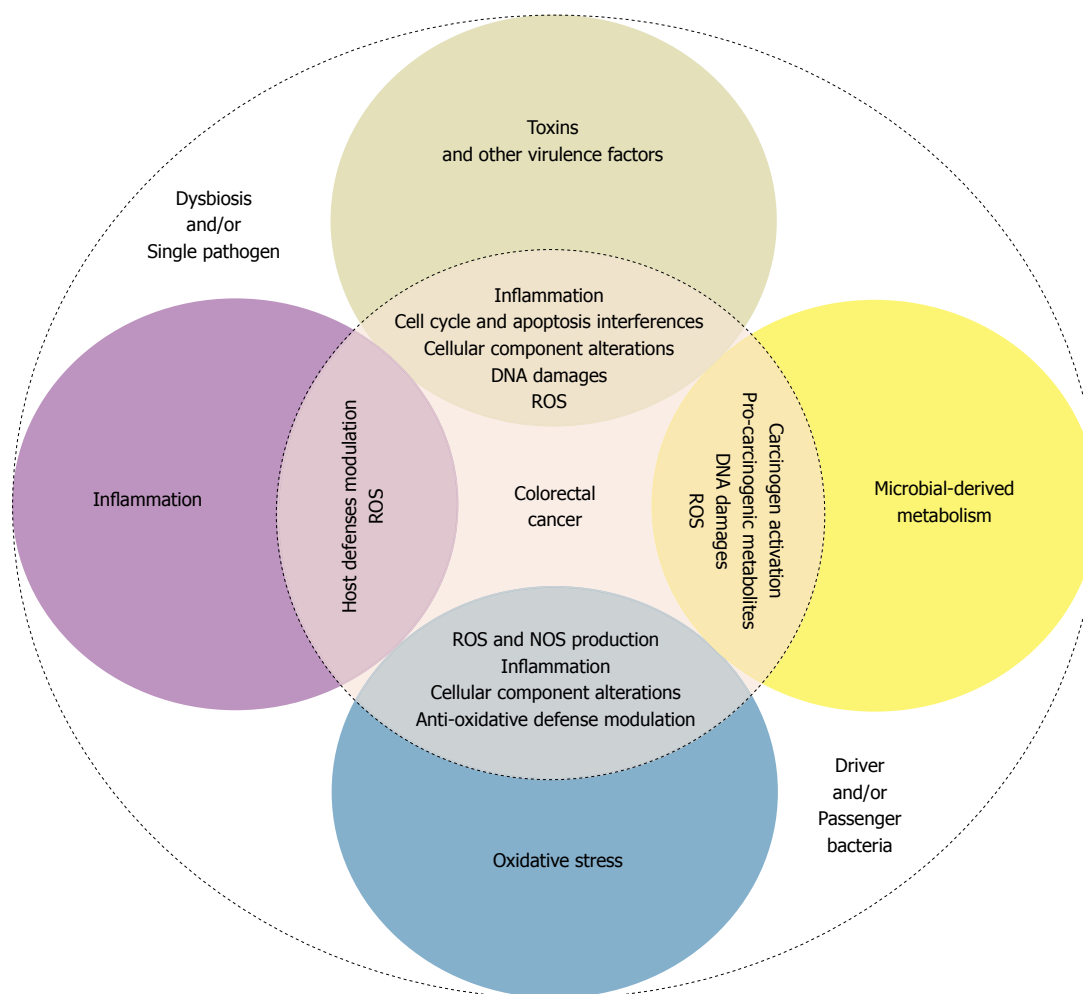


Figure 1 Suspected mechanisms by which the gut bacterial microbiota participates in colorectal carcinogenesis. The bacterial microbiota induces colorectal carcinogenesis through several mechanisms. The primary suspected mechanisms are bacteria-derived genotoxin production, microbial-derived metabolism, the modulation of host defenses and inflammation pathways, oxidative stress induction, and anti-oxidative defense regulation. These mechanisms result in various cellular effects and alterations of host defenses that lead to genomic instability and epithelial cell proliferation, which are involved in colorectal carcinogenesis. ROS: Reactive oxygen species; NOS: Nitrogen species.

primarily in the *Enterobacteriaceae* family, especially in the *E. coli* B2 phylogroup^[115,119]. Interestingly, other micro-organisms, such as *Proteus mirabilis* and *Klebsiella pneumoniae*, which are associated with bacterial-induced colitis and an immunodeficient-CRC-mouse model, also carry the *pks* island^[140-142]. Colibactin induces ROS formation, double-strand DNA breaks, and the associated ATM-mediated DNA damage response, cell cycle arrest and genomic instability^[82,118]. Furthermore, colibactin-producing *E. coli* can enhance tumor growth by inducing the emergence of senescent epithelial cells secreting growth factors and/or by the secretion of pro-tumoral molecules by infiltrating cells^[58,114,120]. Interestingly, even though toxin-producing bacteria constitute a minority of the colonic microbiota, metatranscriptomic analyses performed on human CRC tissues samples reveal a high expression of these toxins in the colon^[143].

Microbial-derived metabolism affecting carcinogenesis

In addition to the involvement of bacterial virulence

factors, such as genotoxin production, it has become increasingly clear that microbial-derived metabolism strongly impacts CRC development^[144]. These metabolic activities may affect colorectal carcinogenesis *via* the following several processes: regulating the generation of CRC-promoting secondary bile acids; the metabolic activation or inactivation of pro-carcinogenic compounds, dietary phytochemicals and xenobiotics; hormone metabolism; and the modification of inflammation pathways^[58].

The interplay between diet, bile acids and gut microbiota is complex. Indeed, high-fat diets are correlated with increased bile secretion and an increased risk of CRC^[145-147]. Primary bile acids excreted into the gut are converted through microbial derived-metabolism, including hydrolase activities, into secondary bile acids. These acids are used by microorganisms as an energy source^[148] but are also known to be involved in many colorectal carcinogenesis-linked process, such as apoptosis, cell proliferation, DNA damage induction and tumor promotion^[27]. Bernstein

et al.^[146] reported a higher incidence of tumors in bile acid-exposed gut in a mouse model. Indeed, bacteria-transformed bile acids can result in DNA damage by the production of pro-oxidative molecules, such as ROS and nitrogen species (NOS)^[149,150]. Therefore, chronic exposure to increased levels of secondary bile acids may favor the induction of DNA damage, leading to genomic instability, which is involved in CRC. Furthermore, strong antimicrobial bile acid activities lead to significant changes in the gut microbiota composition, with a relative increase in some *Gammaproteobacteria* and *Bacteroidetes* species that are associated with CRC^[151].

Some carcinogens are inactivated in the liver by glucuronic acid-mediated conjugation and are excreted *via* bile in the digestive tract. In the gut, and particularly in the colon, this process may be reversed by bacterial β -glucuronidase activity, which can lead to CRC. Indeed, Takada *et al.*^[152] reported a decrease in tumor number with the inhibition of bacterial β -glucuronidase activity in a CRC rat model. Moreover, fecal β -glucuronidase activity is increased in patients with CRC compared to healthy controls^[153]. These results strongly support the involvement of bacterial β -glucuronidase activity in the initiation and progression of CRC *via* the reactivation of toxic components. Moreover, bacterial β -glucuronidase activity plays a major role in the metabolism of xenobiotics and affects the activity and side effects of certain antitumor drugs^[58,154]. For example, irinotecan, a commonly used chemotherapy for CRC that is inactivated in the liver, is locally modified in the gut, generating intermediate molecules without systemic anti-tumor effect and inducing major treatment-limiting side effects (*e.g.*, severe diarrhea). These undesired effects can be prevented with the use of antibiotics or bacterial β -glucuronidase inhibitors^[155].

Contrary to microbial carbohydrate fermentation, which can benefit the host through the generation of short chain fatty acids (*e.g.*, butyrate, acetate, propionate)^[156], microbial protein fermentation generates potentially toxic and pro-carcinogenic metabolites involved in CRC, such as phenols, sulfides, ammonia and nitrosamines^[58]. It has been reported that protein-rich and low-carbohydrate diets can lead to the increased microbial production of toxic metabolites to the detriment of cancer-protective metabolites^[157], increasing the risk for CRC^[158]. However, concentration-dependent effects on CRC have been described for butyrate. At lower concentrations, butyrate appears to stimulate epithelial cell proliferation^[159], while other studies have demonstrated anti-proliferative and anti-cancer properties of this compound^[160]. A subset of bacteria, including *Bacteroidetes* and *Firmicutes* species, produce potentially bioactive substances *via* the degradation of amino acids, especially nitrogenous compounds, in the gut^[161-163]. These compounds can exert carcinogenic effects through DNA alkylation,

leading to mutations that have been reported in Western diet-linked CRC^[161-163]. Moreover, sulfides produced in the gut by the bacterial reduction of dietary sulphate, as well as the metabolism of other compounds^[164], are enterotoxic^[165]. These sulfides have genotoxic effects on human cell lines at physiological concentrations^[166]. These effects occur primarily *via* the induction of ROS formation and DNA damage^[167].

Chronic and/or excessive consumption of alcohol has been found to be an important risk factor for many cancers, including CRC^[168]. Microbial metabolism may contribute to the toxicity of alcohol, especially in the gastrointestinal tract, where aerobic and facultative anaerobic bacteria convert ethanol to acetaldehyde^[169]. Indeed, acetaldehyde is known to be a highly toxic and pro-carcinogenic compound with various negative effects, ranging from DNA damage and impaired DNA excision repair to the degradation of folate. All of these effects have been implicated in colorectal carcinogenesis^[169-171]. The role of microbiota in this process has also been reinforced by Homann *et al.*^[172], who reported that the conversion of ethanol to acetaldehyde was inhibited by the use of antibiotics, such as ciprofloxacin. This drug kills primarily aerobic and facultative anaerobic bacterial populations^[172].

Host defense modulation and inflammation

As previously mentioned, the intestinal mucosa constitute the first line of defense against gut commensal or pathogen bacteria and related microbial molecules. Intestinal epithelial cells need to rapidly detect the presence of pathogens in order to mount a suitable immune response. However, these cells also must maintain a moderate immune response against or tolerance for non-pathogenic bacteria^[173]. The maintenance of gut homeostasis and these interactions between the host and microbiota involve innate immunity receptors, such as Toll Like Receptors (TLRs) and Nod Like Receptors (NLRs), which recognize particular molecular motifs associated with pathogens. The activation of these receptors leads to a cellular response, including the activation of MAPK, NF- κ B or PI3K/AKT signaling pathways^[174]. The activation of these pathways can induce the expression of pro-inflammatory cytokines (*e.g.*, TNF- α , IL-6, IL-8) and/or antimicrobial peptides, all of which are involved in the development of an inflammatory response. Indeed, a decrease in intestinal tumor number has been reported in azoxymethane (AOM)-treated APC^{Min/+} or IL-10^{-/-} mice invalidated for *MyD88* gene, which encodes a TLR signaling adaptor^[175,176]. However, these results remain controversial given that another recent study reported that *MyD88*^{-/-} mice treated with AOM/dextran sulfate sodium (DSS) exhibit increased susceptibility to colorectal tumors. These results suggest that the role of TLR signaling and the host response in CRC differ between colitis-associated and chemically induced CRC^[177]. Moreover, *Nod1*- or *Nod2*-deficient APC^{Min/+}

and AOM/DSS-treated mice exhibit an increase in the number of colorectal tumors compared to control animals, suggesting the involvement of NLRs in colorectal carcinogenesis^[55,178]. These results highlight the major role of the host immune response to gut microbiota in CRC development.

It is now well established that inflammatory bowel disease patients, who are known to have an increased risk for developing CRC, present many changes in their microbiota composition^[179-181]. For these reasons, the involvement of inflammation in the establishment of dysbiosis-related CRC is increasingly evident. Some *in vivo* studies have shown that the gut microbiota composition differs between AOM-treated and untreated *IL-10*^{-/-} mice. Moreover the emergence of dysbiosis after each cycle of DSS treatment in mice treated with AOM has been observed^[82,182]. Taken together, these data constitute strong evidence that inflammation plays an important role in the modulation of the microbiota and dysbiosis emergence during colorectal carcinogenesis.

However, inflammation could also be linked to the host response induced by bacteria during CRC development. It has been previously reported in a CRC-predisposed APC^{Min/+} mouse model that BFT induces a Th17 pro-inflammatory response, which is involved in the development of early-stage tumors^[127]. Furthermore, the contribution of *S. bovis* to colorectal carcinogenesis is associated with the increased expression of pro-inflammatory genes, such as IL-1, IL-8 and COX-2^[90]. Moreover, APC^{Min/+} and *IL-10*^{-/-} mice infected with *F. nucleatum* and *E. faecalis*, respectively, exhibit increased immune cell infiltration in tumors and colonic mucosa and heightened expression of pro-inflammatory cytokines, such as TNF- α , IL-1 β , IL-6 and IL-8^[77,104,183]. *E. coli* is one of the best-characterized bacteria associated with inflammatory bowel disease. An abnormal colonization of the gut mucosa by adherent and invasive *E. coli* in inflammatory bowel disease patients has been reported^[110,111]. Moreover, Raisch *et al.*^[184] showed that CRC-associated *E. coli* can induce the expression of the pro-inflammatory gene COX-2 in macrophages, supporting the bacterial modulation of inflammation in colorectal carcinogenesis.

Oxidative stress and anti-oxidative defenses modulation

ROS induction appears to have a major and central role in microbiota-linked CRC *via* the previously described mechanisms. ROS can be generated by cells during infection and inflammation, as previously discussed, or directly by gut microbiota^[177]. The induction of ROS is known to be a major defense mechanism of infected cells, contributing to the elimination of bacteria^[48,119]. It has been reported in both *in vitro* and *in vivo* studies that some Enterococci species, especially *E. faecalis*, generate hydroxyl radicals^[102,103]. These radicals are powerful mutagens that can cause DNA breaks, point

mutations and protein-DNA crosslinking, all contributing to genomic instability in CRC^[185-187]. Furthermore, this bacterium can induce aneuploidy in colonic epithelial cells, and the use of inhibitors of ROS and NOS can prevent this effect, supporting the role of bacterial-induced oxidative stress in colorectal carcinogenesis^[82]. Moreover, the role of *H. pylori* in gastric carcinogenesis *via* the induction of oxidative stress has been clearly demonstrated^[49], and this species is able to both produce and induce the production of ROS by immune cells. In this manner, *H. pylori* affects many signal transduction pathways in gastric cells and thereby promotes gastric cancer. It could thus extrapolate the fact that existence of colorectal chemical-, bacterial- and/or immune-induced inflammation also induces the recruitment of neutrophils and macrophages, which are major sources of ROS, leading to the genetic and epigenetic alterations involved in CRC^[188-190]. The gut microbiota also promote host-derived production of nitric oxide and its secondary NOS, especially through the activation of macrophages in the inflammation response, which can induce DNA damage. Some bacterial species can directly generate NOS^[191]. Sobko *et al.*^[192,193] reported that *Lactobacilli* and *Bifidobacteria* generate significant levels of NOS in germ-free and monoassociated mice and that a nitrate-enriched diet increased NOS production. However, it is not clear whether in ROS and NOS produced *in vivo* are sufficiently long lived to diffuse from immune cells to the extracellular matrix and subsequently enter the nucleus of epithelial cells to induce damage DNA^[194].

Oxidative stress is defined by an imbalance between the levels of pro-oxidative molecules (*e.g.*, ROS and NOS) and the effectiveness of anti-oxidative defenses^[195]. Oxidative stress results in primarily irreversible direct or organic substrate-mediated cell damage, including DNA breaks and damage, protein aggregation or fragmentation, and cellular membranes dysfunction^[185,195]. The toxic effects of ROS and NOS are thus balanced by various enzymatic and non-enzymatic anti-oxidative defenses, which help to regulate ROS/NOS production and repair mechanisms^[196,197]. DNA repair mechanisms are altered in CRC^[198,199]. Moreover, under specific conditions, the balance between pro- and anti-oxidative compounds is lost, especially in cases of bacterial infection. The bacterial modulation of anti-oxidative defenses, especially DNA repair systems, is becoming increasingly clear. Mangerich *et al.*^[200] reported decreased expression of DNA repair and oxidative response genes in the colons of colitis-induced CRC mice model that were infected with *H. hepaticus*. Moreover, Maddocks *et al.*^[112,121] reported *in vitro* results demonstrating the ability of some enteropathogenic *E. coli* strains to downregulate the DNA MMR system in infected HT-29 intestinal epithelial cells *via* post-transcriptional effect of a secreted effector protein. This downregulation of the MMR system led to the

accumulation of mutations involved in colorectal carcinogenesis^[112,121]. Furthermore, a study on APC^{Min/+} MMR-deficient mice strongly supports a possible role of an interaction between gut microbiota and MMR deficiency in CRC induction^[159]. Indeed, Belcheva *et al.*^[159] reported that (1) altering the microbiota composition reduces tumorigenesis; (2) gut microbes can fuel the hyperproliferation of MMR-altered intestinal epithelial cells; and (3) the MMR pathway has a role in regulating APC/ β -catenin activity and modulating the differentiation of transit-amplifying cells in the colon^[159]. Moreover, Viljoen *et al.*^[109] reported increased colonic colonization by *F. nucleatum* in CRC patients with an MMR deficiency-linked microsatellite instability phenotype. All of these data strengthen the hypothesis of an interaction between the gut microbiota and the DNA repair system in colorectal carcinogenesis.

MICROBIOTA IMBALANCE AND CLINICAL IMPLICATIONS

As previously mentioned, the gut microbiota likely plays a major role in the promotion and progression of CRC *via* several mechanisms, including inflammation, metabolism and genotoxicity. There are therefore many possible ways by which to target the microbiota in terms of CRC prevention strategies. Indeed, the use of probiotics or fecal transplantation protocols could combat CRC-associated dysbiosis and thus restore eubiosis in chronic diseases, helping to reduce microbiota-induced genotoxicity and activation of inflammatory, proliferative and pro-carcinogenic pathways^[58]. However, this microbiota-targeting therapy approach has not been well studied in CRC.

Genotoxins are a target of interest in the context of CRC treatment. For example, supportive evidence has recently been provided that colibactin-producing *E. coli* could be major actors in CRC-related genomic instability^[58,114,115,119,120]. Colibactin synthesis requires the serine enzyme ClbP, which acts as a peptidase to produce colibactin NRPS compounds^[201-203]. On this basis, it has been reported that boronic acid compounds, which are potent inhibitors of active serine enzymes, suppress the genotoxic activity of colibactin-producing *E. coli* *in vitro* and *in vivo*^[204]. In CRC mice model, treatment with such compounds was shown to prevent cell proliferation and genotoxin-induced tumorigenesis compared to water-treated mice^[204].

Changes in gut microbiota composition could also lead to an altered host immune response. On this basis, some authors have studied the impact of the oral administration of probiotics on immunologic signaling. These studies have provided supportive evidence that the gut microbiota plays an essential role in intestinal epigenomic mechanisms of the host^[205,206]. Moreover, it was reported that the deletion of lipoteichoic acid (LTA), a TLR2 ligand, normalizes

innate and adaptive pathogenic immune responses and decreases the number of tumors in a CRC-predisposed murine model. It was also reported that LTA-deficient *Lactobacillus acidophilus* (1) decreased inflammation and protected against CRC^[207]; (2) prevented or induced the regression of established colitis and polyposis^[207,208]; and (3) downregulated downstream signaling^[209], and stimulated tumor suppressor gene expression in CRC cell lines^[210]. Furthermore, some commensal bacteria, such as *Bifidobacterium breve* and *Lactobacillus rhamnosus*, inhibit the production of pro-inflammatory cytokines and decrease host DNA methylation and histone acetylation events that are involved in colorectal carcinogenesis^[211-213]. All of these results underline a feasible microbiota-targeting therapy approach in CRC through the use of probiotics or genetically modified bacteria. Even if fecal transplantation has not been well studied in CRC, future transplantation studies using germ-free CRC animal models likely represent an important next step in this line of inquiry.

To provide adequate treatments as part of modern personalized medicine, accurate prognosis factor have yet to be established. Prognostic factors in CRC primarily depend on morphologic and histologic results. Recently, there has been a significant body of work involving the gut microbiota as a prognosis factor in CRC. Bonnet *et al.*^[108] reported increased colonic colonization by cyclomodulin-producing *E. coli* in advanced stage CRC, that have been ever described by Viljoen *et al.*^[109] for enterotoxigenic *B. fragilis* and *F. nucleatum*. These results support a possible use of microbiota CRC prognosis markers that may be used to improve patient selection for aggressive, suitable treatments.

CONCLUSION

The advent of modern molecular microbiota sequencing techniques has strongly improved the characterization of microbiota variations in CRC. However, a better understanding of the interactions between the host and pathogens in colorectal carcinogenesis requires further microbiota functional studies, especially with respect to metabolomics and RNA sequencing approaches. All of the studies published in this regard have been performed without classifying tumors according to their molecular phenotype. Investigations should also consider the heterogeneity of CRC tumors by studying microbiota imbalances in relation to molecular pathways involved in colorectal carcinogenesis, such as chromosomal and microsatellite instabilities or CpG island methylator phenotypes. In summary, the role of the microbiota in CRC is increasingly evident and perhaps represents a new approach towards the improved therapeutic management of patients with CRC.

REFERENCES

- 1 **Savage DC.** Microbial ecology of the gastrointestinal tract. *Annu Rev Microbiol* 1977; **31**: 107-133 [PMID: 334036 DOI: 10.1146/annurev.mi.31.100177.000543]
- 2 **Suau A, Bonnet R, Sutren M, Godon JJ, Gibson GR, Collins MD, Doré J.** Direct analysis of genes encoding 16S rRNA from complex communities reveals many novel molecular species within the human gut. *Appl Environ Microbiol* 1999; **65**: 4799-4807 [PMID: 10543789]
- 3 **Neish AS.** Microbes in gastrointestinal health and disease. *Gastroenterology* 2009; **136**: 65-80 [PMID: 19026645 DOI: 10.1053/j.gastro.2008.10.080]
- 4 **Goncharova GI, Dorofeichuk VG, Smolianskaia AZ, Sokolova KlA.** [Microbial ecology of the intestines in health and in pathology]. *Antibiot Khimioter* 1989; **34**: 462-466 [PMID: 2802880]
- 5 **Dominguez-Bello MG, Blaser MJ, Ley RE, Knight R.** Development of the human gastrointestinal microbiota and insights from high-throughput sequencing. *Gastroenterology* 2011; **140**: 1713-1719 [PMID: 21530737 DOI: 10.1053/j.gastro.2011.02.011]
- 6 **Mulder IE, Schmidt B, Lewis M, Delday M, Stokes CR, Bailey M, Aminov RI, Gill BP, Pluske JR, Mayer CD, Kelly D.** Restricting microbial exposure in early life negates the immune benefits associated with gut colonization in environments of high microbial diversity. *PLoS One* 2011; **6**: e28279 [PMID: 22216092 DOI: 10.1371/journal.pone.0028279]
- 7 **Dethlefsen L, Eckburg PB, Bik EM, Relman DA.** Assembly of the human intestinal microbiota. *Trends Ecol Evol* 2006; **21**: 517-523 [PMID: 16820245 DOI: 10.1016/j.tree.2006.06.013]
- 8 **Stanghellini V, Barbara G, Cremon C, Cogliandro R, Antonucci A, Gabusi V, Frisoni C, De Giorgio R, Grasso V, Serra M, Corinaldesi R.** Gut microbiota and related diseases: clinical features. *Intern Emerg Med* 2010; **5** Suppl 1: S57-S63 [PMID: 20865476 DOI: 10.1007/s11739-010-0451-0]
- 9 **Claesson MJ, Cusack S, O'Sullivan O, Greene-Diniz R, de Weerd H, Flannery E, Marchesi JR, Falush D, Dinan T, Fitzgerald G, Stanton C, van Sinderen D, O'Connor M, Harnedy N, O'Connor K, Henry C, O'Mahony D, Fitzgerald AP, Shanahan F, Twomey C, Hill C, Ross RP, O'Toole PW.** Composition, variability, and temporal stability of the intestinal microbiota of the elderly. *Proc Natl Acad Sci USA* 2011; **108** Suppl 1: 4586-4591 [PMID: 20571116 DOI: 10.1073/pnas.1000097107]
- 10 **Rajilić-Stojanović M, Heilig HG, Molenaar D, Kajander K, Surakka A, Smidt H, de Vos WM.** Development and application of the human intestinal tract chip, a phylogenetic microarray: analysis of universally conserved phylotypes in the abundant microbiota of young and elderly adults. *Environ Microbiol* 2009; **11**: 1736-1751 [PMID: 19508560 DOI: 10.1111/j.1462-2920.2009.01900.x]
- 11 **Turnbaugh PJ, Backhed F, Fulton L, Gordon JI.** Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. *Cell Host Microbe* 2008; **3**: 213-223 [PMID: 18407065 DOI: 10.1016/j.chom.2008.02.015]
- 12 **Zwiehler J, Liszt K, Handschur M, Lassl C, Lapin A, Haslberger AG.** Combined PCR-DGGE fingerprinting and quantitative-PCR indicates shifts in fecal population sizes and diversity of Bacteroides, bifidobacteria and Clostridium cluster IV in institutionalized elderly. *Exp Gerontol* 2009; **44**: 440-446 [PMID: 19376217 DOI: 10.1016/j.exger.2009.04.002]
- 13 **Ivanov II, Frutos Rde L, Manel N, Yoshinaga K, Rifkin DB, Sartor RB, Finlay BB, Littman DR.** Specific microbiota direct the differentiation of IL-17-producing T-helper cells in the mucosa of the small intestine. *Cell Host Microbe* 2008; **4**: 337-349 [PMID: 18854238 DOI: 10.1016/j.chom.2008.09.009]
- 14 **Marchesi JR.** Human distal gut microbiome. *Environ Microbiol* 2011; **13**: 3088-3102 [PMID: 21906225 DOI: 10.1111/j.1462-2920.2011.02574.x]
- 15 **Hakansson A, Molin G.** Gut microbiota and inflammation. *Nutrients* 2011; **3**: 637-682 [PMID: 22254115 DOI: 10.3390/nu3060637]
- 16 **Sekirov I, Russell SL, Antunes LC, Finlay BB.** Gut microbiota in health and disease. *Physiol Rev* 2010; **90**: 859-904 [PMID: 20664075 DOI: 10.1152/physrev.00045.2009]
- 17 **Kelly D, Mulder IE.** Microbiome and immunological interactions. *Nutr Rev* 2012; **70** Suppl 1: S18-S30 [PMID: 22861803 DOI: 10.1111/j.1753-4887.2012.00498.x]
- 18 **Chen W, Liu F, Ling Z, Tong X, Xiang C.** Human intestinal lumen and mucosa-associated microbiota in patients with colorectal cancer. *PLoS One* 2012; **7**: e39743 [PMID: 22761885 DOI: 10.1371/journal.pone.0039743]
- 19 **Arthur JC, Jobin C.** The struggle within: microbial influences on colorectal cancer. *Inflamm Bowel Dis* 2011; **17**: 396-409 [PMID: 20848537 DOI: 10.1002/ibd.21354]
- 20 **Ley RE, Backhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI.** Obesity alters gut microbial ecology. *Proc Natl Acad Sci USA* 2005; **102**: 11070-11075 [PMID: 16033867 DOI: 10.1073/pnas.0504978102]
- 21 **Mai V.** Dietary modification of the intestinal microbiota. *Nutr Rev* 2004; **62**: 235-242 [PMID: 15291396 DOI: 10.1111/j.1753-4887.2004.tb00045.x]
- 22 **Rastall RA.** Bacteria in the gut: friends and foes and how to alter the balance. *J Nutr* 2004; **134**: 2022S-2026S [PMID: 15284393]
- 23 **Hopkins MJ, Sharp R, Macfarlane GT.** Age and disease related changes in intestinal bacterial populations assessed by cell culture, 16S rRNA abundance, and community cellular fatty acid profiles. *Gut* 2001; **48**: 198-205 [PMID: 11156640 DOI: 10.1136/gut.48.2.198]
- 24 **Zotendal EG, Akkermans ADL, Akkermans-van Vilet WM, de Visser JAGM, de Vos WM.** The host genotype affects the bacterial community in the human gastrointestinal tract. *Microb Ecol Health Dis* 2001; **13**: 129-134 [DOI: 10.1080/089106001750462669]
- 25 **Tlaskalová-Hogenová H, Stepanková R, Hudcovic T, Tucková L, Cukrowska B, Lodinová-Zádníková R, Kozáková H, Rossmann P, Bártová J, Sokol D, Funda DP, Borovská D, Reháková Z, Sinkora J, Hofman J, Drastich P, Kokesová A.** Commensal bacteria (normal microflora), mucosal immunity and chronic inflammatory and autoimmune diseases. *Immunol Lett* 2004; **93**: 97-108 [PMID: 15158604 DOI: 10.1016/j.imlet.2004.02.005]
- 26 **Peterson DA, Frank DN, Pace NR, Gordon JI.** Metagenomic approaches for defining the pathogenesis of inflammatory bowel diseases. *Cell Host Microbe* 2008; **3**: 417-427 [PMID: 18541218 DOI: 10.1016/j.chom.2008.05.001]
- 27 **Boleij A, Tjalsma H.** Gut bacteria in health and disease: a survey on the interface between intestinal microbiology and colorectal cancer. *Biol Rev Camb Philos Soc* 2012; **87**: 701-730 [PMID: 22296522 DOI: 10.1111/j.1469-185X.2012.00218.x]
- 28 **O'Hara AM, Shanahan F.** The gut flora as a forgotten organ. *EMBO Rep* 2006; **7**: 688-693 [PMID: 16819463 DOI: 10.1038/sj.embor.7400731]
- 29 **Umesaki Y, Okada Y, Matsumoto S, Imaoka A, Setoyama H.** Segmented filamentous bacteria are indigenous intestinal bacteria that activate intraepithelial lymphocytes and induce MHC class II molecules and fucosyl asialo GMI glycolipids on the small intestinal epithelial cells in the ex-germ-free mouse. *Microbiol Immunol* 1995; **39**: 555-562 [PMID: 7494493]
- 30 **Xu J, Gordon JI.** Honor thy symbionts. *Proc Natl Acad Sci USA* 2003; **100**: 10452-10459 [PMID: 12923294 DOI: 10.1073/pnas.1734063100]
- 31 **Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL.** An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* 2005; **122**: 107-118 [PMID: 16009137 DOI: 10.1016/j.cell.2005.05.007]
- 32 **Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R.** Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* 2004; **118**: 229-241 [PMID: 15260992 DOI: 10.1016/j.cell.2004.07.002]
- 33 **Weinstein PD, Cebra JJ.** The preference for switching to IgA expression by Peyer's patch germinal center B cells is likely due to the intrinsic influence of their microenvironment. *J Immunol* 1991; **147**: 4126-4135 [PMID: 1753088]
- 34 **Cebra JJ.** Influences of microbiota on intestinal immune system development. *Am J Clin Nutr* 1999; **69**: 1046S-1051S [PMID: 20664075 DOI: 10.1152/physrev.00045.2009]

- 10232647]
- 35 **Shanahan F.** The host-microbe interface within the gut. *Best Pract Res Clin Gastroenterol* 2002; **16**: 915-931 [PMID: 12473298 DOI: 10.1053/bega.2002.0342]
 - 36 **Li Y, Kundu P, Seow SW, de Matos CT, Aronsson L, Chin KC, Kärre K, Pettersson S, Greicius G.** Gut microbiota accelerate tumor growth via c-jun and STAT3 phosphorylation in APCMin/+ mice. *Carcinogenesis* 2012; **33**: 1231-1238 [PMID: 22461519 DOI: 10.1093/carcin/bgs137]
 - 37 **Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, Semenkovich CF, Gordon JI.** The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* 2004; **101**: 15718-15723 [PMID: 15505215 DOI: 10.1073/pnas.0407076101]
 - 38 **Bäckhed F, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI.** Host-bacterial mutualism in the human intestine. *Science* 2005; **307**: 1915-1920 [PMID: 15790844 DOI: 10.1126/science.1104816]
 - 39 **Laparra JM, Sanz Y.** Interactions of gut microbiota with functional food components and nutraceuticals. *Pharmacol Res* 2010; **61**: 219-225 [PMID: 19914380 DOI: 10.1016/j.phrs.2009.11.001]
 - 40 **Wall R, Ross RP, Shanahan F, O'Mahony L, O'Mahony C, Coakley M, Hart O, Lawlor P, Quigley EM, Kiely B, Fitzgerald GF, Stanton C.** Metabolic activity of the enteric microbiota influences the fatty acid composition of murine and porcine liver and adipose tissues. *Am J Clin Nutr* 2009; **89**: 1393-1401 [PMID: 19357220 DOI: 10.3945/ajcn.2008.27023]
 - 41 **Manning TS, Gibson GR.** Microbial-gut interactions in health and disease. Prebiotics. *Best Pract Res Clin Gastroenterol* 2004; **18**: 287-298 [PMID: 15123070 DOI: 10.1016/j.bpg.2003.10.008]
 - 42 **Simmering R, Pforte H, Jacobasch G, Blaut M.** The growth of the flavonoid-degrading intestinal bacterium, *Eubacterium ramulus*, is stimulated by dietary flavonoids in vivo. *FEMS Microbiol Ecol* 2002; **40**: 243-248 [PMID: 19709232 DOI: 10.1111/j.1574-6941.2002.tb00957.x]
 - 43 **Stecher B, Hardt WD.** The role of microbiota in infectious disease. *Trends Microbiol* 2008; **16**: 107-114 [PMID: 18280160 DOI: 10.1016/j.tim.2007.12.008]
 - 44 **Alexander DD, Cushing CA, Lowe KA, Scurman B, Roberts MA.** Meta-analysis of animal fat or animal protein intake and colorectal cancer. *Am J Clin Nutr* 2009; **89**: 1402-1409 [PMID: 19261724 DOI: 10.3945/ajcn.2008.26838]
 - 45 **Sandler RS.** Epidemiology and risk factors for colorectal cancer. *Gastroenterol Clin North Am* 1996; **25**: 717-735 [PMID: 8960889 DOI: 10.1016/S0889-8553(05)70271-5]
 - 46 **Knudson A.** Alfred Knudson and his two-hit hypothesis. (Interview by Ezzie Hutchinson). *Lancet Oncol* 2001; **2**: 642-645 [PMID: 11902557 DOI: 10.1016/S1470-2045(01)00524-1]
 - 47 **Zur Hausen H.** The search for infectious causes of human cancers: where and why. *Virology* 2009; **392**: 1-10 [PMID: 19720205 DOI: 10.1016/j.virol.2009.06.001]
 - 48 **Collins D, Hogan AM, Winter DC.** Microbial and viral pathogens in colorectal cancer. *Lancet Oncol* 2011; **12**: 504-512 [PMID: 21067973 DOI: 10.1016/S1470-2045(10)70186-8]
 - 49 **Handa O, Naito Y, Yoshikawa T.** *Helicobacter pylori*: a ROS-inducing bacterial species in the stomach. *Inflamm Res* 2010; **59**: 997-1003 [PMID: 20820854 DOI: 10.1007/s00011-010-0245-x]
 - 50 **Sears CL, Garrett WS.** Microbes, microbiota, and colon cancer. *Cell Host Microbe* 2014; **15**: 317-328 [PMID: 24629338 DOI: 10.1016/j.chom.2014.02.007]
 - 51 **Proctor LM.** The Human Microbiome Project in 2011 and beyond. *Cell Host Microbe* 2011; **10**: 287-291 [PMID: 22018227 DOI: 10.1016/j.chom.2011.10.001]
 - 52 **Weisburger JH, Reddy BS, Narisawa T, Wynder EL.** Germ-free status and colon tumor induction by N-methyl-N'-nitro-N-nitrosoguanidine. *Proc Soc Exp Biol Med* 1975; **148**: 1119-1121 [PMID: 1129327]
 - 53 **Vannucci L, Stepankova R, Kozakova H, Fiserova A, Rossmann P, Tlaskalova-Hogenova H.** Colorectal carcinogenesis in germ-free and conventionally reared rats: different intestinal environments affect the systemic immunity. *Int J Oncol* 2008; **32**: 609-617 [PMID: 18292938]
 - 54 **Tjalsma H, Boleij A, Marchesi JR, Dutilh BE.** A bacterial driver-passenger model for colorectal cancer: beyond the usual suspects. *Nat Rev Microbiol* 2012; **10**: 575-582 [PMID: 22728587 DOI: 10.1038/nrmicro2819]
 - 55 **Couturier-Maillard A, Secher T, Rehman A, Normand S, De Arcangelis A, Haesler R, Huot L, Grandjean T, Bressenot A, Delanoye-Crespin A, Gaillot O, Schreiber S, Lemoine Y, Ryffel B, Hot D, Nuñez G, Chen G, Rosenstiel P, Chamaillard M.** NOD2-mediated dysbiosis predisposes mice to transmissible colitis and colorectal cancer. *J Clin Invest* 2013; **123**: 700-711 [PMID: 23281400 DOI: 10.1172/JCI62236]
 - 56 **Hu B, Elinav E, Huber S, Strowig T, Hao L, Hafemann A, Jin C, Wunderlich C, Wunderlich T, Eisenbarth SC, Flavell RA.** Microbiota-induced activation of epithelial IL-6 signaling links inflammasome-driven inflammation with transmissible cancer. *Proc Natl Acad Sci USA* 2013; **110**: 9862-9867 [PMID: 23696660 DOI: 10.1073/pnas.1307575110]
 - 57 **Human Microbiome Project Consortium.** Structure, function and diversity of the healthy human microbiome. *Nature* 2012; **486**: 207-214 [PMID: 22699609 DOI: 10.1038/nature11234]
 - 58 **Schwabe RF, Jobin C.** The microbiome and cancer. *Nat Rev Cancer* 2013; **13**: 800-812 [PMID: 24132111 DOI: 10.1038/nrc3610]
 - 59 **Hajishengallis G, Darveau RP, Curtis MA.** The keystone-pathogen hypothesis. *Nat Rev Microbiol* 2012; **10**: 717-725 [PMID: 22941505 DOI: 10.1038/nrmicro2873]
 - 60 **Sanapareddy N, Legge RM, Jovov B, McCoy A, Burcal L, Araujo-Perez F, Randall TA, Galanko J, Benson A, Sandler RS, Rawls JF, Abdo Z, Fodor AA, Keku TO.** Increased rectal microbial richness is associated with the presence of colorectal adenomas in humans. *ISME J* 2012; **6**: 1858-1868 [PMID: 22622349 DOI: 10.1038/ismej.2012.43]
 - 61 **Sobhani I, Tap J, Roudot-Thoraval F, Roperch JP, Letulle S, Langella P, Corthier G, Tran Van Nhieu J, Furet JP.** Microbial dysbiosis in colorectal cancer (CRC) patients. *PLoS One* 2011; **6**: e16393 [PMID: 21297998 DOI: 10.1371/journal.pone.0016393]
 - 62 **Wang T, Cai G, Qiu Y, Fei N, Zhang M, Pang X, Jia W, Cai S, Zhao L.** Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. *ISME J* 2012; **6**: 320-329 [PMID: 21850056 DOI: 10.1038/ismej.2011.109]
 - 63 **Ahn J, Sinha R, Pei Z, Dominianni C, Wu J, Shi J, Goedert JJ, Hayes RB, Yang L.** Human gut microbiome and risk for colorectal cancer. *J Natl Cancer Inst* 2013; **105**: 1907-1911 [PMID: 24316595 DOI: 10.1093/jnci/djt300]
 - 64 **Wu N, Yang X, Zhang R, Li J, Xiao X, Hu Y, Chen Y, Yang F, Lu N, Wang Z, Luan C, Liu Y, Wang B, Xiang C, Wang Y, Zhao F, Gao GF, Wang S, Li L, Zhang H, Zhu B.** Dysbiosis signature of fecal microbiota in colorectal cancer patients. *Microb Ecol* 2013; **66**: 462-470 [PMID: 23733170 DOI: 10.1007/s00248-013-0245-9]
 - 65 **Abdulamir AS, Hafidh RR, Abu Bakar F.** The association of *Streptococcus bovis/galloyticus* with colorectal tumors: the nature and the underlying mechanisms of its etiological role. *J Exp Clin Cancer Res* 2011; **30**: 11 [PMID: 21247505 DOI: 10.1186/1756-9966-30-11]
 - 66 **Klein RS, Recco RA, Catalano MT, Edberg SC, Casey JI, Steigbigel NH.** Association of *Streptococcus bovis* with carcinoma of the colon. *N Engl J Med* 1977; **297**: 800-802 [PMID: 408687 DOI: 10.1056/NEJM197710132971503]
 - 67 **Grahn N, Hmani-Aifa M, Fransén K, Söderkvist P, Monstein HJ.** Molecular identification of *Helicobacter DNA* present in human colorectal adenocarcinomas by 16S rDNA PCR amplification and pyrosequencing analysis. *J Med Microbiol* 2005; **54**: 1031-1035 [PMID: 16192433 DOI: 10.1099/jmm.0.46122-0]
 - 68 **Jones M, Helliwell P, Pritchard C, Tharakan J, Mathew J.** *Helicobacter pylori* in colorectal neoplasms: is there an aetiological relationship? *World J Surg Oncol* 2007; **5**: 51 [PMID: 17498313 DOI: 10.1186/1477-7819-5-51]
 - 69 **Zumkeller N, Brenner H, Zwahlen M, Rothenbacher D.** *Helicobacter pylori* infection and colorectal cancer risk: a meta-analysis. *Helicobacter* 2006; **11**: 75-80 [PMID: 16579836 DOI: 10.1111/j.1523-5378.2006.00381.x]

- 70 **Housseau F**, Sears CL. Enterotoxigenic *Bacteroides fragilis* (ETBF)-mediated colitis in Min (Apc^{+/-}) mice: a human commensal-based murine model of colon carcinogenesis. *Cell Cycle* 2010; **9**: 3-5 [PMID: 20009569]
- 71 **Toprak NU**, Yagci A, Gulluoglu BM, Akin ML, Demirkalem P, Celenk T, Soyletir G. A possible role of *Bacteroides fragilis* enterotoxin in the aetiology of colorectal cancer. *Clin Microbiol Infect* 2006; **12**: 782-786 [PMID: 16842574 DOI: 10.1111/j.1469-0691.2006.01494.x]
- 72 **Wu S**, Morin PJ, Maouyo D, Sears CL. *Bacteroides fragilis* enterotoxin induces c-Myc expression and cellular proliferation. *Gastroenterology* 2003; **124**: 392-400 [PMID: 12557145 DOI: 10.1053/gast.2003.50047]
- 73 **Balamurugan R**, Rajendiran E, George S, Samuel GV, Ramakrishna BS. Real-time polymerase chain reaction quantification of specific butyrate-producing bacteria, *Desulfovibrio* and *Enterococcus faecalis* in the feces of patients with colorectal cancer. *J Gastroenterol Hepatol* 2008; **23**: 1298-1303 [PMID: 18624900 DOI: 10.1111/j.1440-1746.2008.05490.x]
- 74 **Chew SS**, Lubowski DZ. *Clostridium septicum* and malignancy. *ANZ J Surg* 2001; **71**: 647-649 [PMID: 11736823]
- 75 **Hermesen JL**, Schurr MJ, Kudsk KA, Faucher LD. Phenotyping *Clostridium septicum* infection: a surgeon's infectious disease. *J Surg Res* 2008; **148**: 67-76 [PMID: 18570933 DOI: 10.1016/j.jss.2008.02.027]
- 76 **Mirza NN**, McCloud JM, Cheetham MJ. *Clostridium septicum* sepsis and colorectal cancer - a reminder. *World J Surg Oncol* 2009; **7**: 73 [PMID: 19807912 DOI: 10.1186/1477-7819-7-73]
- 77 **Kostic AD**, Chun E, Robertson L, Glickman JN, Gallini CA, Michaud M, Clancy TE, Chung DC, Lochhead P, Hold GL, El-Omar EM, Brenner D, Fuchs CS, Meyerson M, Garrett WS. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. *Cell Host Microbe* 2013; **14**: 207-215 [PMID: 23954159 DOI: 10.1016/j.chom.2013.07.007]
- 78 **McCoy AN**, Araújo-Pérez F, Azcárate-Peril A, Yeh JJ, Sandler RS, Keku TO. *Fusobacterium* is associated with colorectal adenomas. *PLoS One* 2013; **8**: e53653 [PMID: 23335968 DOI: 10.1371/journal.pone.0053653]
- 79 **Rubinstein MR**, Wang X, Liu W, Hao Y, Cai G, Han YW. *Fusobacterium nucleatum* promotes colorectal carcinogenesis by modulating E-cadherin/ β -catenin signaling via its FadA adhesin. *Cell Host Microbe* 2013; **14**: 195-206 [PMID: 23954158 DOI: 10.1016/j.chom.2013.07.012]
- 80 **Martin HM**, Campbell BJ, Hart CA, Mpofu C, Nayar M, Singh R, Englyst H, Williams HF, Rhodes JM. Enhanced *Escherichia coli* adherence and invasion in Crohn's disease and colon cancer. *Gastroenterology* 2004; **127**: 80-93 [PMID: 15236175 DOI: 10.1053/j.gastro.2004.03.054]
- 81 **Swidsinski A**, Khilkin M, Kerjaschki D, Schreiber S, Ortner M, Weber J, Lochs H. Association between intraepithelial *Escherichia coli* and colorectal cancer. *Gastroenterology* 1998; **115**: 281-286 [PMID: 9679033 DOI: 10.1016/S0016-5085(98)70194-5]
- 82 **Arthur JC**, Perez-Chanona E, Mühlbauer M, Tomkovich S, Uronis JM, Fan TJ, Campbell BJ, Abujamel T, Dogan B, Rogers AB, Rhodes JM, Stintzi A, Simpson KW, Hansen JJ, Keku TO, Fodor AA, Jobin C. Intestinal inflammation targets cancer-inducing activity of the microbiota. *Science* 2012; **338**: 120-123 [PMID: 22903521 DOI: 10.1126/science.1224820]
- 83 **McCoy WC**, Mason JM 3rd. Enterococcal endocarditis associated with carcinoma of the sigmoid; report of a case. *J Med Assoc State Ala* 1951; **21**: 162-166 [PMID: 14880846]
- 84 **Hoppes WL**, Lerner PI. Nonenterococcal group-D streptococcal endocarditis caused by *Streptococcus bovis*. *Ann Intern Med* 1974; **81**: 588-593 [PMID: 4422602 DOI: 10.7326/0003-4819-81-5-588]
- 85 **Klein RS**, Catalano MT, Edberg SC, Casey JI, Steigbigel NH. *Streptococcus bovis* septicemia and carcinoma of the colon. *Ann Intern Med* 1979; **91**: 560-562 [PMID: 484953 DOI: 10.7326/0003-4819-91-4-560]
- 86 **Dubrow R**, Edberg S, Wikfors E, Callan D, Troncale F, Vender R, Brand M, Yapp R. Fecal carriage of *Streptococcus bovis* and colorectal adenomas. *Gastroenterology* 1991; **101**: 721-725 [PMID: 1823534]
- 87 **Potter MA**, Cunliffe NA, Smith M, Miles RS, Flapan AD, Dunlop MG. A prospective controlled study of the association of *Streptococcus bovis* with colorectal carcinoma. *J Clin Pathol* 1998; **51**: 473-474 [PMID: 9771449 DOI: 10.1136/jcp.51.6.473]
- 88 **Boleij A**, van Gelder MM, Swinkels DW, Tjalsma H. Clinical Importance of *Streptococcus gallolyticus* infection among colorectal cancer patients: systematic review and meta-analysis. *Clin Infect Dis* 2011; **53**: 870-878 [PMID: 21960713 DOI: 10.1093/cid/cir609]
- 89 **Zoetendal EG**, von Wright A, Vilpponen-Salmela T, Ben-Amor K, Akkermans AD, de Vos WM. Mucosa-associated bacteria in the human gastrointestinal tract are uniformly distributed along the colon and differ from the community recovered from feces. *Appl Environ Microbiol* 2002; **68**: 3401-3407 [PMID: 12089021]
- 90 **Abdulmir AS**, Hafidh RR, Bakar FA. Molecular detection, quantification, and isolation of *Streptococcus gallolyticus* bacteria colonizing colorectal tumors: inflammation-driven potential of carcinogenesis via IL-1, COX-2, and IL-8. *Mol Cancer* 2010; **9**: 249 [PMID: 20846456 DOI: 10.1186/1476-4598-9-249]
- 91 **Boleij A**, Tjalsma H. The itinerary of *Streptococcus gallolyticus* infection in patients with colonic malignant disease. *Lancet Infect Dis* 2013; **13**: 719-724 [PMID: 23831427 DOI: 10.1016/S1473-3099(13)70107-5]
- 92 **Guo Y**, Li HY. Association between *Helicobacter pylori* infection and colorectal neoplasm risk: a meta-analysis based on East Asian population. *J Cancer Res Ther* 2014; **10** Suppl: 263-266 [PMID: 25693932 DOI: 10.4103/0973-1482.151482]
- 93 **Higashi H**, Tsutsumi R, Fujita A, Yamazaki S, Asaka M, Azuma T, Hatakeyama M. Biological activity of the *Helicobacter pylori* virulence factor CagA is determined by variation in the tyrosine phosphorylation sites. *Proc Natl Acad Sci USA* 2002; **99**: 14428-14433 [PMID: 12391297 DOI: 10.1073/pnas.222375399]
- 94 **Shmueli H**, Passaro D, Figer A, Niv Y, Pitlik S, Samra Z, Koren R, Yahav J. Relationship between *Helicobacter pylori* CagA status and colorectal cancer. *Am J Gastroenterol* 2001; **96**: 3406-3410 [PMID: 11774957 DOI: 10.1111/j.1572-0241.2001.05342.x]
- 95 **Huang JY**, Lee SM, Mazmanian SK. The human commensal *Bacteroides fragilis* binds intestinal mucin. *Anaerobe* 2011; **17**: 137-141 [PMID: 21664470 DOI: 10.1016/j.anaerobe.2011.05.017]
- 96 **Macfarlane S**, Woodmansey EJ, Macfarlane GT. Colonization of mucin by human intestinal bacteria and establishment of biofilm communities in a two-stage continuous culture system. *Appl Environ Microbiol* 2005; **71**: 7483-7492 [PMID: 16269790 DOI: 10.1128/AEM.71.11.7483-7492.2005]
- 97 **Sears CL**. Enterotoxigenic *Bacteroides fragilis*: a rogue among symbiotes. *Clin Microbiol Rev* 2009; **22**: 349-69, Table of Contents [PMID: 19366918 DOI: 10.1128/CMR.00053-08]
- 98 **Zitomersky NL**, Coyne MJ, Comstock LE. Longitudinal analysis of the prevalence, maintenance, and IgA response to species of the order Bacteroidales in the human gut. *Infect Immun* 2011; **79**: 2012-2020 [PMID: 21402766 DOI: 10.1128/IAI.01348-10]
- 99 **Boleij A**, Hechenbleikner EM, Goodwin AC, Badani R, Stein EM, Lazarev MG, Ellis B, Carroll KC, Albesiano E, Wick EC, Platz EA, Pardoll DM, Sears CL. The *Bacteroides fragilis* toxin gene is prevalent in the colon mucosa of colorectal cancer patients. *Clin Infect Dis* 2015; **60**: 208-215 [PMID: 25305284 DOI: 10.1093/cid/ciu787]
- 100 **Rhee KJ**, Wu S, Wu X, Huso DL, Karim B, Franco AA, Rabizadeh S, Golub JE, Mathews LE, Shin J, Sartor RB, Golenbock D, Hamad AR, Gan CM, Housseau F, Sears CL. Induction of persistent colitis by a human commensal, enterotoxigenic *Bacteroides fragilis*, in wild-type C57BL/6 mice. *Infect Immun* 2009; **77**: 1708-1718 [PMID: 19188353 DOI: 10.1128/IAI.00814-08]
- 101 **Pillar CM**, Gilmore MS. Enterococcal virulence--pathogenicity island of *E. faecalis*. *Front Biosci* 2004; **9**: 2335-2346 [PMID: 15353291 DOI: 10.2741/1400]
- 102 **Huycke MM**, Moore D, Joyce W, Wise P, Shepard L, Kotake Y, Gilmore MS. Extracellular superoxide production by *Enterococcus faecalis* requires demethylmenaquinone and is attenuated by

- functional terminal quinol oxidases. *Mol Microbiol* 2001; **42**: 729-740 [PMID: 11722738 DOI: 10.1046/j.1365-2958.2001.02638.x]
- 103 **Huycke MM**, Moore DR. In vivo production of hydroxyl radical by *Enterococcus faecalis* colonizing the intestinal tract using aromatic hydroxylation. *Free Radic Biol Med* 2002; **33**: 818-826 [PMID: 12208369 DOI: 10.1016/S0891-5849(02)00977-2]
- 104 **Balish E**, Warner T. *Enterococcus faecalis* induces inflammatory bowel disease in interleukin-10 knockout mice. *Am J Pathol* 2002; **160**: 2253-2257 [PMID: 12057927 DOI: 10.1016/S0002-9440(10)61172-8]
- 105 **Dylewski J**, Luteran L. Septic arthritis and *Clostridium septicum*: a clue to colon cancer. *CMAJ* 2010; **182**: 1446-1447 [PMID: 20855487 DOI: 10.1503/cmaj.091946]
- 106 **Kostic AD**, Gevers D, Pedamallu CS, Michaud M, Duke F, Earl AM, Ojesina AI, Jung J, Bass AJ, Taberero J, Baselga J, Liu C, Shivdasani RA, Ogino S, Birren BW, Huttenhower C, Garrett WS, Meyerson M. Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. *Genome Res* 2012; **22**: 292-298 [PMID: 22009990 DOI: 10.1101/gr.126573.111]
- 107 **Marchesi JR**, Dutilh BE, Hall N, Peters WH, Roelofs R, Boleij A, Tjalsma H. Towards the human colorectal cancer microbiome. *PLoS One* 2011; **6**: e20447 [PMID: 21647227 DOI: 10.1371/journal.pone.0020447]
- 108 **Bonnet M**, Buc E, Sauvanet P, Darcha C, Dubois D, Pereira B, Déchelotte P, Bonnet R, Pezet D, Darfeuille-Michaud A. Colonization of the human gut by *E. coli* and colorectal cancer risk. *Clin Cancer Res* 2014; **20**: 859-867 [PMID: 24334760 DOI: 10.1158/1078-0432.CCR-13-1343]
- 109 **Viljoen KS**, Dakshinamurthy A, Goldberg P, Blackburn JM. Quantitative profiling of colorectal cancer-associated bacteria reveals associations between *Fusobacterium* spp., enterotoxigenic *Bacteroides fragilis* (ETBF) and clinicopathological features of colorectal cancer. *PLoS One* 2015; **10**: e0119462 [PMID: 25751261 DOI: 10.1371/journal.pone.0119462]
- 110 **Darfeuille-Michaud A**, Boudeau J, Bulois P, Neut C, Glasser AL, Barnich N, Bringer MA, Swidsinski A, Beaugerie L, Colombel JF. High prevalence of adherent-invasive *Escherichia coli* associated with ileal mucosa in Crohn's disease. *Gastroenterology* 2004; **127**: 412-421 [PMID: 15300573 DOI: 10.1053/j.gastro.2004.04.061]
- 111 **Darfeuille-Michaud A**, Neut C, Barnich N, Lederman E, Di Martino P, Desreumaux P, Gambiez L, Joly B, Cortot A, Colombel JF. Presence of adherent *Escherichia coli* strains in ileal mucosa of patients with Crohn's disease. *Gastroenterology* 1998; **115**: 1405-1413 [PMID: 9834268 DOI: 10.1016/S0016-5085(98)70019-8]
- 112 **Maddocks OD**, Short AJ, Sonnenberg MS, Bader S, Harrison DJ. Attaching and effacing *Escherichia coli* downregulate DNA mismatch repair protein in vitro and are associated with colorectal adenocarcinomas in humans. *PLoS One* 2009; **4**: e5517 [PMID: 19436735 DOI: 10.1371/journal.pone.0005517]
- 113 **Martins SA**, Prazeres DM, Cabral JM, Monteiro GA. Comparison of real-time polymerase chain reaction and hybridization assays for the detection of *Escherichia coli* genomic DNA in process samples and pharmaceutical-grade plasmid DNA products. *Anal Biochem* 2003; **322**: 127-129 [PMID: 14705789 DOI: 10.1016/j.ab.2003.07.004]
- 114 **Arthur JC**, Jobin C. The complex interplay between inflammation, the microbiota and colorectal cancer. *Gut Microbes* 2013; **4**: 253-258 [PMID: 23549517 DOI: 10.4161/gmic.24220]
- 115 **Buc E**, Dubois D, Sauvanet P, Raisch J, Delmas J, Darfeuille-Michaud A, Pezet D, Bonnet R. High prevalence of mucosa-associated *E. coli* producing cyclomodulin and genotoxin in colon cancer. *PLoS One* 2013; **8**: e56964 [PMID: 23457644 DOI: 10.1371/journal.pone.0056964]
- 116 **Prorok-Hamon M**, Friswell MK, Alswied A, Roberts CL, Song F, Flanagan PK, Knight P, Codling C, Marchesi JR, Winstanley C, Hall N, Rhodes JM, Campbell BJ. Colonic mucosa-associated diffusely adherent afaC+ *Escherichia coli* expressing lpfA and pks are increased in inflammatory bowel disease and colon cancer. *Gut* 2014; **63**: 761-770 [PMID: 23846483 DOI: 10.1136/gutjnl-2013-304739]
- 117 **Raisch J**, Buc E, Bonnet M, Sauvanet P, Vazeille E, de Vallée A, Déchelotte P, Darcha C, Pezet D, Bonnet R, Bringer MA, Darfeuille-Michaud A. Colon cancer-associated B2 *Escherichia coli* colonize gut mucosa and promote cell proliferation. *World J Gastroenterol* 2014; **20**: 6560-6572 [PMID: 24914378 DOI: 10.3748/wjg.v20.i21.6560]
- 118 **Cuevas-Ramos G**, Petit CR, Marcq I, Boury M, Oswald E, Nougayrède JP. *Escherichia coli* induces DNA damage in vivo and triggers genomic instability in mammalian cells. *Proc Natl Acad Sci USA* 2010; **107**: 11537-11542 [PMID: 20534522 DOI: 10.1073/pnas.1001261107]
- 119 **Nougayrède JP**, Homburg S, Taieb F, Boury M, Brzuszkiewicz E, Gottschalk G, Buchrieser C, Hacker J, Dobrindt U, Oswald E. *Escherichia coli* induces DNA double-strand breaks in eukaryotic cells. *Science* 2006; **313**: 848-851 [PMID: 16902142 DOI: 10.1126/science.1127059]
- 120 **Cougoux A**, Dalmasso G, Martinez R, Buc E, Delmas J, Gibold L, Sauvanet P, Darcha C, Déchelotte P, Bonnet M, Pezet D, Wodrich H, Darfeuille-Michaud A, Bonnet R. Bacterial genotoxin colibactin promotes colon tumour growth by inducing a senescence-associated secretory phenotype. *Gut* 2014; **63**: 1932-1942 [PMID: 24658599 DOI: 10.1136/gutjnl-2013-305257]
- 121 **Maddocks OD**, Scanlon KM, Sonnenberg MS. An *Escherichia coli* effector protein promotes host mutation via depletion of DNA mismatch repair proteins. *MBio* 2013; **4**: e00152-e00113 [PMID: 23781066 DOI: 10.1128/mBio.00152-13]
- 122 **Escobar-Páramo P**, Grenet K, Le Menac'h A, Rode L, Salgado E, Amorin C, Gouriou S, Picard B, Rahimy MC, Andremont A, Denamur E, Ruimy R. Large-scale population structure of human commensal *Escherichia coli* isolates. *Appl Environ Microbiol* 2004; **70**: 5698-5700 [PMID: 15345464 DOI: 10.1128/AEM.70.9.5698-5700.2004]
- 123 **Le Gall T**, Clermont O, Gouriou S, Picard B, Nassif X, Denamur E, Tenaillon O. Extraintestinal virulence is a coincidental by-product of commensalism in B2 phylogenetic group *Escherichia coli* strains. *Mol Biol Evol* 2007; **24**: 2373-2384 [PMID: 17709333 DOI: 10.1093/molbev/msm172]
- 124 **Han YW**, Ikegami A, Rajanna C, Kawsar HI, Zhou Y, Li M, Sojar HT, Genco RJ, Kuramitsu HK, Deng CX. Identification and characterization of a novel adhesin unique to oral *Fusobacterium*. *J Bacteriol* 2005; **187**: 5330-5340 [PMID: 16030227 DOI: 10.1128/JB.187.15.5330-5340.2005]
- 125 **Fox JG**, Wang TC. Inflammation, atrophy, and gastric cancer. *J Clin Invest* 2007; **117**: 60-69 [PMID: 17200707 DOI: 10.1172/JCI30111]
- 126 **Ohnishi N**, Yuasa H, Tanaka S, Sawa H, Miura M, Matsui A, Higashi H, Musashi M, Iwabuchi K, Suzuki M, Yamada G, Azuma T, Hatakeyama M. Transgenic expression of *Helicobacter pylori* CagA induces gastrointestinal and hematopoietic neoplasms in mouse. *Proc Natl Acad Sci USA* 2008; **105**: 1003-1008 [PMID: 18192401 DOI: 10.1073/pnas.0711183105]
- 127 **Wu S**, Rhee KJ, Albesiano E, Rabizadeh S, Wu X, Yen HR, Huso DL, Brancati FL, Wick E, McAllister F, Housseau F, Pardoll DM, Sears CL. A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. *Nat Med* 2009; **15**: 1016-1022 [PMID: 19701202 DOI: 10.1038/nm.2015]
- 128 **Moncrief JS**, Obiso R, Barroso LA, Kling JJ, Wright RL, Van Tassel RL, Lysterly DM, Wilkins TD. The enterotoxin of *Bacteroides fragilis* is a metalloprotease. *Infect Immun* 1995; **63**: 175-181 [PMID: 7806355]
- 129 **Goodwin AC**, Destefano Shields CE, Wu S, Huso DL, Wu X, Murray-Stewart TR, Hacker-Prietz A, Rabizadeh S, Woster PM, Sears CL, Casero RA. Polyamine catabolism contributes to enterotoxigenic *Bacteroides fragilis*-induced colon tumorigenesis. *Proc Natl Acad Sci USA* 2011; **108**: 15354-15359 [PMID: 21876161 DOI: 10.1073/pnas.1010203108]
- 130 **Wu S**, Lim KC, Huang J, Saidi RF, Sears CL. *Bacteroides fragilis* enterotoxin cleaves the zonula adherens protein, E-cadherin. *Proc Natl Acad Sci USA* 1998; **95**: 14979-14984 [PMID: 9844001 DOI: 10.1073/pnas.95.25.14979]

- 131 **Wu S**, Powell J, Mathioudakis N, Kane S, Fernandez E, Sears CL. Bacteroides fragilis enterotoxin induces intestinal epithelial cell secretion of interleukin-8 through mitogen-activated protein kinases and a tyrosine kinase-regulated nuclear factor-kappaB pathway. *Infect Immun* 2004; **72**: 5832-5839 [PMID: 15385484 DOI: 10.1128/IAI.72.10.5832-5839.2004]
- 132 **Wu S**, Shin J, Zhang G, Cohen M, Franco A, Sears CL. The Bacteroides fragilis toxin binds to a specific intestinal epithelial cell receptor. *Infect Immun* 2006; **74**: 5382-5390 [PMID: 16926433 DOI: 10.1128/IAI.00060-06]
- 133 **Nesić D**, Hsu Y, Stebbins CE. Assembly and function of a bacterial genotoxin. *Nature* 2004; **429**: 429-433 [PMID: 15164065 DOI: 10.1038/nature02532]
- 134 **Nougayrède JP**, Taieb F, De Rycke J, Oswald E. Cyclomodulins: bacterial effectors that modulate the eukaryotic cell cycle. *Trends Microbiol* 2005; **13**: 103-110 [PMID: 15737728 DOI: 10.1016/j.tim.2005.01.002]
- 135 **Oswald E**, Nougayrède JP, Taieb F, Sugai M. Bacterial toxins that modulate host cell-cycle progression. *Curr Opin Microbiol* 2005; **8**: 83-91 [PMID: 15694861 DOI: 10.1016/j.mib.2004.12.011]
- 136 **Travaglione S**, Fabbri A, Fiorentini C. The Rho-activating CNF1 toxin from pathogenic E. coli: a risk factor for human cancer development? *Infect Agent Cancer* 2008; **3**: 4 [PMID: 18336718 DOI: 10.1186/1750-9378-3-4]
- 137 **Smith JL**, Bayles DO. The contribution of cytolethal distending toxin to bacterial pathogenesis. *Crit Rev Microbiol* 2006; **32**: 227-248 [PMID: 17123907 DOI: 10.1080/10408410601023557]
- 138 **Ge Z**, Rogers AB, Feng Y, Lee A, Xu S, Taylor NS, Fox JG. Bacterial cytolethal distending toxin promotes the development of dysplasia in a model of microbially induced hepatocarcinogenesis. *Cell Microbiol* 2007; **9**: 2070-2080 [PMID: 17441986 DOI: 10.1111/j.1462-5822.2007.00939.x]
- 139 **Ge Z**, Schauer DB, Fox JG. In vivo virulence properties of bacterial cytolethal-distending toxin. *Cell Microbiol* 2008; **10**: 1599-1607 [PMID: 18489725 DOI: 10.1111/j.1462-5822.2008.01173.x]
- 140 **Garrett WS**, Gallini CA, Yatsunenkov T, Michaud M, DuBois A, Delaney ML, Punit S, Karlsson M, Bry L, Glickman JN, Gordon JL, Onderdonk AB, Glimcher LH. Enterobacteriaceae act in concert with the gut microbiota to induce spontaneous and maternally transmitted colitis. *Cell Host Microbe* 2010; **8**: 292-300 [PMID: 20833380 DOI: 10.1016/j.chom.2010.08.004]
- 141 **Garrett WS**, Punit S, Gallini CA, Michaud M, Zhang D, Sigrist KS, Lord GM, Glickman JN, Glimcher LH. Colitis-associated colorectal cancer driven by T-bet deficiency in dendritic cells. *Cancer Cell* 2009; **16**: 208-219 [PMID: 19732721 DOI: 10.1016/j.ccr.2009.07.015]
- 142 **Putze J**, Hennequin C, Nougayrède JP, Zhang W, Homburg S, Karch H, Bringer MA, Fayolle C, Carniel E, Rabsch W, Oelschlaeger TA, Oswald E, Forestier C, Hacker J, Dobrindt U. Genetic structure and distribution of the colibactin genomic island among members of the family Enterobacteriaceae. *Infect Immun* 2009; **77**: 4696-4703 [PMID: 19720753 DOI: 10.1128/IAI.00522-09]
- 143 **Dutilh BE**, Backus L, van Hijum SA, Tjalsma H. Screening metatranscriptomes for toxin genes as functional drivers of human colorectal cancer. *Best Pract Res Clin Gastroenterol* 2013; **27**: 85-99 [PMID: 23768555 DOI: 10.1016/j.bpg.2013.03.008]
- 144 **Louis P**, Hold GL, Flint HJ. The gut microbiota, bacterial metabolites and colorectal cancer. *Nat Rev Microbiol* 2014; **12**: 661-672 [PMID: 25198138 DOI: 10.1038/nrmicro3344]
- 145 **Barrasa JI**, Olmo N, Lizarbe MA, Turnay J. Bile acids in the colon, from healthy to cytotoxic molecules. *Toxicol In Vitro* 2013; **27**: 964-977 [PMID: 23274766 DOI: 10.1016/j.tiv.2012.12.020]
- 146 **Bernstein H**, Bernstein C, Payne CM, Dvorak K. Bile acids as endogenous etiologic agents in gastrointestinal cancer. *World J Gastroenterol* 2009; **15**: 3329-3340 [PMID: 19610133]
- 147 **Ou J**, DeLany JP, Zhang M, Sharma S, O'Keefe SJ. Association between low colonic short-chain fatty acids and high bile acids in high colon cancer risk populations. *Nutr Cancer* 2012; **64**: 34-40 [PMID: 22136517 DOI: 10.1080/01635581.2012.630164]
- 148 **Philipp B**. Bacterial degradation of bile salts. *Appl Microbiol Biotechnol* 2011; **89**: 903-915 [PMID: 21088832 DOI: 10.1007/s00253-010-2998-0]
- 149 **Bernstein H**, Bernstein C, Payne CM, Dvorakova K, Garewal H. Bile acids as carcinogens in human gastrointestinal cancers. *Mutat Res* 2005; **589**: 47-65 [PMID: 15652226 DOI: 10.1016/j.mrrev.2004.08.001]
- 150 **Dvorak K**, Payne CM, Chavarria M, Ramsey L, Dvorakova B, Bernstein H, Holubec H, Sampliner RE, Guy N, Condon A, Bernstein C, Green SB, Prasad A, Garewal HS. Bile acids in combination with low pH induce oxidative stress and oxidative DNA damage: relevance to the pathogenesis of Barrett's oesophagus. *Gut* 2007; **56**: 763-771 [PMID: 17145738 DOI: 10.1136/gut.2006.103697]
- 151 **Yoshimoto S**, Loo TM, Atarashi K, Kanda H, Sato S, Oyadomari S, Iwakura Y, Oshima K, Morita H, Hattori M, Honda K, Ishikawa Y, Hara E, Ohtani N. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature* 2013; **499**: 97-101 [PMID: 23803760 DOI: 10.1038/nature12347]
- 152 **Takada H**, Hirooka T, Hiramatsu Y, Yamamoto M. Effect of beta-glucuronidase inhibitor on azoxymethane-induced colonic carcinogenesis in rats. *Cancer Res* 1982; **42**: 331-334 [PMID: 7053860]
- 153 **Kim DH**, Jin YH. Intestinal bacterial beta-glucuronidase activity of patients with colon cancer. *Arch Pharm Res* 2001; **24**: 564-567 [PMID: 11794536]
- 154 **Haiser HJ**, Turnbaugh PJ. Is it time for a metagenomic basis of therapeutics? *Science* 2012; **336**: 1253-1255 [PMID: 22674325 DOI: 10.1126/science.1224396]
- 155 **Wallace BD**, Wang H, Lane KT, Scott JE, Orans J, Koo JS, Venkatesh M, Jobin C, Yeh LA, Mani S, Redinbo MR. Alleviating cancer drug toxicity by inhibiting a bacterial enzyme. *Science* 2010; **330**: 831-835 [PMID: 21051639 DOI: 10.1126/science.1191175]
- 156 **Nyangale EP**, Mottram DS, Gibson GR. Gut microbial activity, implications for health and disease: the potential role of metabolite analysis. *J Proteome Res* 2012; **11**: 5573-5585 [PMID: 23116228 DOI: 10.1021/pr300637d]
- 157 **Russell WR**, Gratz SW, Duncan SH, Holtrop G, Ince J, Scobbie L, Duncan G, Johnstone AM, Lobley GE, Wallace RJ, Duthie GG, Flint HJ. High-protein, reduced-carbohydrate weight-loss diets promote metabolite profiles likely to be detrimental to colonic health. *Am J Clin Nutr* 2011; **93**: 1062-1072 [PMID: 21389180 DOI: 10.3945/ajcn.110.002188]
- 158 **Hughes R**, Cross AJ, Pollock JR, Bingham S. Dose-dependent effect of dietary meat on endogenous colonic N-nitrosation. *Carcinogenesis* 2001; **22**: 199-202 [PMID: 11159760]
- 159 **Belcheva A**, Irrazabal T, Robertson SJ, Streutker C, Maughan H, Rubino S, Moriyama EH, Copeland JK, Kumar S, Green B, Geddes K, Pezo RC, Navarre WW, Milosevic M, Wilson BC, Girardin SE, Wolever TM, Edelmann W, Guttman DS, Philpott DJ, Martin A. Gut microbial metabolism drives transformation of MSH2-deficient colon epithelial cells. *Cell* 2014; **158**: 288-299 [PMID: 25036629 DOI: 10.1016/j.cell.2014.04.051]
- 160 **Singh N**, Gurav A, Sivaprakasam S, Brady E, Padia R, Shi H, Thangaraju M, Prasad PD, Manicassamy S, Munn DH, Lee JR, Offermanns S, Ganapathy V. Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. *Immunity* 2014; **40**: 128-139 [PMID: 24412617 DOI: 10.1016/j.immuni.2013.12.007]
- 161 **Gill CI**, Rowland IR. Diet and cancer: assessing the risk. *Br J Nutr* 2002; **88** Suppl 1: S73-S87 [PMID: 12215186 DOI: 10.1079/BJN2002632]
- 162 **Russell WR**, Duncan SH, Scobbie L, Duncan G, Cantlay L, Calder AG, Anderson SE, Flint HJ. Major phenylpropanoid-derived metabolites in the human gut can arise from microbial fermentation of protein. *Mol Nutr Food Res* 2013; **57**: 523-535 [PMID: 23349065 DOI: 10.1002/mnfr.201200594]
- 163 **Loh YH**, Jakszyn P, Luben RN, Mulligan AA, Mitrou PN, Khaw KT. N-Nitroso compounds and cancer incidence: the European Prospective Investigation into Cancer and Nutrition (EPIC)-Norfolk Study. *Am J Clin Nutr* 2011; **93**: 1053-1061 [PMID: 21430112 DOI: 10.3945/ajcn.110.002188]

- 10.3945/ajcn.111.012377]
- 164 **Magée EA**, Richardson CJ, Hughes R, Cummings JH. Contribution of dietary protein to sulfide production in the large intestine: an in vitro and a controlled feeding study in humans. *Am J Clin Nutr* 2000; **72**: 1488-1494 [PMID: 11101476]
- 165 **Roediger WE**, Moore J, Babidge W. Colonic sulfide in pathogenesis and treatment of ulcerative colitis. *Dig Dis Sci* 1997; **42**: 1571-1579 [PMID: 9286219]
- 166 **Attene-Ramos MS**, Nava GM, Muellner MG, Wagner ED, Plewa MJ, Gaskins HR. DNA damage and toxicogenomic analyses of hydrogen sulfide in human intestinal epithelial FHs 74 Int cells. *Environ Mol Mutagen* 2010; **51**: 304-314 [PMID: 20120018 DOI: 10.1002/em.20546]
- 167 **Attene-Ramos MS**, Wagner ED, Gaskins HR, Plewa MJ. Hydrogen sulfide induces direct radical-associated DNA damage. *Mol Cancer Res* 2007; **5**: 455-459 [PMID: 17475672 DOI: 10.1158/1541-7786.MCR-06-0439]
- 168 **Seitz HK**, Stickel F. Molecular mechanisms of alcohol-mediated carcinogenesis. *Nat Rev Cancer* 2007; **7**: 599-612 [PMID: 17646865 DOI: 10.1038/nrc2191]
- 169 **Homann N**. Alcohol and upper gastrointestinal tract cancer: the role of local acetaldehyde production. *Addict Biol* 2001; **6**: 309-323 [PMID: 11900609 DOI: 10.1080/13556210020077028]
- 170 **Hooper SJ**, Wilson MJ, Crean SJ. Exploring the link between microorganisms and oral cancer: a systematic review of the literature. *Head Neck* 2009; **31**: 1228-1239 [PMID: 19475550 DOI: 10.1002/hed.21140]
- 171 **Choi SW**, Kim YI, Weitzel JN, Mason JB. Folate depletion impairs DNA excision repair in the colon of the rat. *Gut* 1998; **43**: 93-99 [PMID: 9771411]
- 172 **Homann N**, Tillonen J, Salaspuro M. Microbially produced acetaldehyde from ethanol may increase the risk of colon cancer via folate deficiency. *Int J Cancer* 2000; **86**: 169-173 [PMID: 10738242]
- 173 **Cario E**. Bacterial interactions with cells of the intestinal mucosa: Toll-like receptors and NOD2. *Gut* 2005; **54**: 1182-1193 [PMID: 15840688 DOI: 10.1136/gut.2004.062794]
- 174 **Karin M**, Greten FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol* 2005; **5**: 749-759 [PMID: 16175180 DOI: 10.1038/nri1703]
- 175 **Rakoff-Nahoum S**, Medzhitov R. Regulation of spontaneous intestinal tumorigenesis through the adaptor protein MyD88. *Science* 2007; **317**: 124-127 [PMID: 17615359 DOI: 10.1126/science.1140488]
- 176 **Uronis JM**, Mühlbauer M, Herfarth HH, Rubinas TC, Jones GS, Jobin C. Modulation of the intestinal microbiota alters colitis-associated colorectal cancer susceptibility. *PLoS One* 2009; **4**: e6026 [PMID: 19551144 DOI: 10.1371/journal.pone.0006026]
- 177 **Irrazábal T**, Belcheva A, Girardin SE, Martin A, Philpott DJ. The multifaceted role of the intestinal microbiota in colon cancer. *Mol Cell* 2014; **54**: 309-320 [PMID: 24766895 DOI: 10.1016/j.molcel.2014.03.039]
- 178 **Chen GY**, Shaw MH, Redondo G, Núñez G. The innate immune receptor Nod1 protects the intestine from inflammation-induced tumorigenesis. *Cancer Res* 2008; **68**: 10060-10067 [PMID: 19074871 DOI: 10.1158/0008-5472.CAN-08-2061]
- 179 **Chassaing B**, Darfeuille-Michaud A. The commensal microbiota and enteropathogens in the pathogenesis of inflammatory bowel diseases. *Gastroenterology* 2011; **140**: 1720-1728 [PMID: 21530738 DOI: 10.1053/j.gastro.2011.01.054]
- 180 **Cunningham D**, Atkin W, Lenz HJ, Lynch HT, Minsky B, Nordlinger B, Starling N. Colorectal cancer. *Lancet* 2010; **375**: 1030-1047 [PMID: 20304247 DOI: 10.1016/S0140-6736(10)60353-4]
- 181 **Manichanh C**, Borruel N, Casellas F, Guarner F. The gut microbiota in IBD. *Nat Rev Gastroenterol Hepatol* 2012; **9**: 599-608 [PMID: 22907164 DOI: 10.1038/nrgastro.2012.152]
- 182 **Zackular JP**, Baxter NT, Iverson KD, Sadler WD, Petrosino JF, Chen GY, Schloss PD. The gut microbiome modulates colon tumorigenesis. *MBio* 2013; **4**: e00692-e00613 [PMID: 24194538 DOI: 10.1128/mBio.00692-13]
- 183 **Yang Y**, Wang X, Moore DR, Lightfoot SA, Huycke MM. TNF- α mediates macrophage-induced bystander effects through Netrin-1. *Cancer Res* 2012; **72**: 5219-5229 [PMID: 22915753 DOI: 10.1158/0008-5472.CAN-12-1463]
- 184 **Raisch J**, Rolhion N, Dubois A, Darfeuille-Michaud A, Bringer MA. Intracellular colon cancer-associated *Escherichia coli* promote protumoral activities of human macrophages by inducing sustained COX-2 expression. *Lab Invest* 2015; **95**: 296-307 [PMID: 25545478 DOI: 10.1038/labinvest.2014.161]
- 185 **Cooke MS**, Evans MD, Dizdaroglu M, Lunec J. Oxidative DNA damage: mechanisms, mutation, and disease. *FASEB J* 2003; **17**: 1195-1214 [PMID: 12832285 DOI: 10.1096/fj.02-0752rev]
- 186 **Evans MD**, Dizdaroglu M, Cooke MS. Oxidative DNA damage and disease: induction, repair and significance. *Mutat Res* 2004; **567**: 1-61 [PMID: 15341901 DOI: 10.1016/j.mrrev.2003.11.001]
- 187 **Wang D**, Kreuzer DA, Essigmann JM. Mutagenicity and repair of oxidative DNA damage: insights from studies using defined lesions. *Mutat Res* 1998; **400**: 99-115 [PMID: 9685598 DOI: 10.1016/S0027-5107(98)00066-9]
- 188 **Bartsch H**, Nair J. Potential role of lipid peroxidation derived DNA damage in human colon carcinogenesis: studies on exocyclic base adducts as stable oxidative stress markers. *Cancer Detect Prev* 2002; **26**: 308-312 [PMID: 12430635 DOI: 10.1016/S0361-090X(02)00093-4]
- 189 **Keshavarzian A**, Zapeda D, List T, Mobarhan S. High levels of reactive oxygen metabolites in colon cancer tissue: analysis by chemiluminescence probe. *Nutr Cancer* 1992; **17**: 243-249 [PMID: 1331990 DOI: 10.1080/01635589209514193]
- 190 **Roessner A**, Kuester D, Malfertheiner P, Schneider-Stock R. Oxidative stress in ulcerative colitis-associated carcinogenesis. *Pathol Res Pract* 2008; **204**: 511-524 [PMID: 18571874 DOI: 10.1016/j.prp.2008.04.011]
- 191 **Lundberg JO**, Weitzberg E, Cole JA, Benjamin N. Nitrate, bacteria and human health. *Nat Rev Microbiol* 2004; **2**: 593-602 [PMID: 15197394 DOI: 10.1038/nrmicro929]
- 192 **Sobko T**, Huang L, Midtvedt T, Norin E, Gustafsson LE, Norman M, Jansson EA, Lundberg JO. Generation of NO by probiotic bacteria in the gastrointestinal tract. *Free Radic Biol Med* 2006; **41**: 985-991 [PMID: 16934682 DOI: 10.1016/j.freeradbiomed.2006.06.020]
- 193 **Sobko T**, Reinders CI, Jansson E, Norin E, Midtvedt T, Lundberg JO. Gastrointestinal bacteria generate nitric oxide from nitrate and nitrite. *Nitric Oxide* 2005; **13**: 272-278 [PMID: 16183308 DOI: 10.1016/j.niox.2005.08.002]
- 194 **Grivannikov SI**, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010; **140**: 883-899 [PMID: 20303878 DOI: 10.1016/j.cell.2010.01.025]
- 195 **Klaunig JE**, Kamendulis LM, Hocevar BA. Oxidative stress and oxidative damage in carcinogenesis. *Toxicol Pathol* 2010; **38**: 96-109 [PMID: 20019356 DOI: 10.1177/0192623309356453]
- 196 **Dröge W**. Free radicals in the physiological control of cell function. *Physiol Rev* 2002; **82**: 47-95 [PMID: 11773609 DOI: 10.1152/physrev.00018.2001]
- 197 **Hazane-Puch F**, Bonnet M, Valenti K, Schnebert S, Kurfurst R, Favier A, Sauvaigo S. Study of fibroblast gene expression in response to oxidative stress induced by hydrogen peroxide or UVA with skin aging. *Eur J Dermatol* 2010; **20**: 308-320 [PMID: 20299309 DOI: 10.1684/ejd.2010.0934]
- 198 **Moreno V**, Gemignani F, Landi S, Gioia-Patricola L, Chabrier A, Blanco I, González S, Guino E, Capellà G, Canzian F. Polymorphisms in genes of nucleotide and base excision repair: risk and prognosis of colorectal cancer. *Clin Cancer Res* 2006; **12**: 2101-2108 [PMID: 16609022 DOI: 10.1158/1078-0432.CCR-05-1363]
- 199 **Peltomäki P**. Role of DNA mismatch repair defects in the pathogenesis of human cancer. *J Clin Oncol* 2003; **21**: 1174-1179 [PMID: 12637487]
- 200 **Mangerich A**, Knutson CG, Parry NM, Muthupalani S, Ye W, Prestwich E, Cui L, McFaline JL, Mobley M, Ge Z, Taghizadeh K, Wishnok JS, Wogan GN, Fox JG, Tannenbaum SR, Dedon PC. Infection-induced colitis in mice causes dynamic and tissue-specific changes in stress response and DNA damage leading to colon

- cancer. *Proc Natl Acad Sci USA* 2012; **109**: E1820-E1829 [PMID: 22689960 DOI: 10.1073/pnas.1207829109]
- 201 **Cougnoux A**, Gibold L, Robin F, Dubois D, Pradel N, Darfeuille-Michaud A, Dalmaso G, Delmas J, Bonnet R. Analysis of structure-function relationships in the colibactin-maturating enzyme ClbP. *J Mol Biol* 2012; **424**: 203-214 [PMID: 23041299 DOI: 10.1016/j.jmb.2012.09.017]
- 202 **Dubois D**, Baron O, Cougnoux A, Delmas J, Pradel N, Boury M, Bouchon B, Bringer MA, Nougayrède JP, Oswald E, Bonnet R. ClbP is a prototype of a peptidase subgroup involved in biosynthesis of nonribosomal peptides. *J Biol Chem* 2011; **286**: 35562-35570 [PMID: 21795676 DOI: 10.1074/jbc.M111.221960]
- 203 **Reimer D**, Pos KM, Thines M, Grün P, Bode HB. A natural prodrug activation mechanism in nonribosomal peptide synthesis. *Nat Chem Biol* 2011; **7**: 888-890 [PMID: 21926994 DOI: 10.1038/nchembio.688]
- 204 **Cougnoux A**, Delmas J, Gibold L, Faïs T, Romagnoli C, Robin F, Cuevas-Ramos G, Oswald E, Darfeuille-Michaud A, Prati F, Dalmaso G, Bonnet R. Small-molecule inhibitors prevent the genotoxic and protumoural effects induced by colibactin-producing bacteria. *Gut* 2015; Epub ahead of print [PMID: 25588406 DOI: 10.1136/gutjnl-2014-307241]
- 205 **Licciardi PV**, Wong SS, Tang ML, Karagiannis TC. Epigenome targeting by probiotic metabolites. *Gut Pathog* 2010; **2**: 24 [PMID: 21172038 DOI: 10.1186/1757-4749-2-24]
- 206 **Ritchie ML**, Romanuk TN. A meta-analysis of probiotic efficacy for gastrointestinal diseases. *PLoS One* 2012; **7**: e34938 [PMID: 22529959 DOI: 10.1371/journal.pone.0034938]
- 207 **Khazaie K**, Zadeh M, Khan MW, Bere P, Gounari F, Dennis K, Blatner NR, Owen JL, Klaenhammer TR, Mohamadzadeh M. Abating colon cancer polyposis by *Lactobacillus acidophilus* deficient in lipoteichoic acid. *Proc Natl Acad Sci USA* 2012; **109**: 10462-10467 [PMID: 22689992 DOI: 10.1073/pnas.1207230109]
- 208 **Mohamadzadeh M**, Pfeiler EA, Brown JB, Zadeh M, Gramarossa M, Managlia E, Bere P, Sarraj B, Khan MW, Pakanati KC, Ansari MJ, O'Flaherty S, Barrett T, Klaenhammer TR. Regulation of induced colonic inflammation by *Lactobacillus acidophilus* deficient in lipoteichoic acid. *Proc Natl Acad Sci USA* 2011; **108** Suppl 1: 4623-4630 [PMID: 21282652 DOI: 10.1073/pnas.1005066107]
- 209 **Saber R**, Zadeh M, Pakanati KC, Bere P, Klaenhammer T, Mohamadzadeh M. Lipoteichoic acid-deficient *Lactobacillus acidophilus* regulates downstream signals. *Immunotherapy* 2011; **3**: 337-347 [PMID: 21395377 DOI: 10.2217/imt.10.119]
- 210 **Lightfoot YL**, Yang T, Sahay B, Mohamadzadeh M. Targeting aberrant colon cancer-specific DNA methylation with lipoteichoic acid-deficient *Lactobacillus acidophilus*. *Gut Microbes* 2013; **4**: 84-88 [PMID: 23137966 DOI: 10.4161/gmic.22822]
- 211 **Ghadimi D**, Helwig U, Schrezenmeir J, Heller KJ, de Vrese M. Epigenetic imprinting by commensal probiotics inhibits the IL-23/IL-17 axis in an in vitro model of the intestinal mucosal immune system. *J Leukoc Biol* 2012; **92**: 895-911 [PMID: 22730546 DOI: 10.1189/jlb.0611286]
- 212 **Hedin C**, Whelan K, Lindsay JO. Evidence for the use of probiotics and prebiotics in inflammatory bowel disease: a review of clinical trials. *Proc Nutr Soc* 2007; **66**: 307-315 [PMID: 17637082 DOI: 10.1017/S0029665107005563]
- 213 **Marteau PR**, de Vrese M, Cellier CJ, Schrezenmeir J. Protection from gastrointestinal diseases with the use of probiotics. *Am J Clin Nutr* 2001; **73**: 430S-436S [PMID: 11157353]
- 214 **Feng Q**, Liang S, Jia H, Stadlmayr A, Tang L, Lan Z, Zhang D, Xia H, Xu X, Jie Z, Su L, Li X, Li X, Li J, Xiao L, Huber-Schönauer U, Niederseer D, Xu X, Al-Aama JY, Yang H, Wang J, Kristiansen K, Arumugam M, Tilg H, Datz C, Wang J. Gut microbiome development along the colorectal adenoma-carcinoma sequence. *Nat Commun* 2015; **6**: 6528 [PMID: 25758642 DOI: 10.1038/ncomms7528]

P- Reviewer: Altomare DF **S- Editor:** Gong ZM **L- Editor:** A
E- Editor: Wang CH





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>



ISSN 1007-9327



9 771007 932045