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## Inactivation of *Bacillus sporothermodurans* LTIS27 spores by high hydrostatic pressure and moderate heat studied by response surface methodology

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### ABSTRACT

*Bacillus sporothermodurans* is a milk spoilage bacterium producing resistant endospores that survive ultra-high temperature treatment. The inactivation of *B. sporothermodurans* LTIS27 spores by combined hydrostatic pressure and heat treatments was studied. A central composite experimental design was used to evaluate the effect of pressure (300–500 MPa), temperature (30–50  C), and pressure-holding time (10–30 min) on the inactivation of spores in distilled water and skim milk. The inactivation observed was shown to fit well with the values predicted by the quadratic equation, since  $R^2_{adj}$  were 0.970 and 0.977 in distilled water and milk, respectively. By analyzing the response surface plots, the inactivation was shown to be higher in distilled water than in milk under all the conditions tested. This was probably due to a protective effect of milk against inactivation by pressure. The optimum process parameter values for a 5-log cycle reduction of spores were calculated as 477 MPa/48  C for 26 min and 495 MPa/49  C for 30 min in water and in milk, respectively. This study shows the efficiency of hydrostatic pressure in combination with moderate temperature to inactivate *B. sporothermodurans* spores. Such treatments could be applied by the dairy industry to ensure the commercial sterility of UHT milk.

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### 1. Introduction

Aerobic spore-forming bacteria of the genus *Bacillus* are often present in raw milk and play an important role in the spoilage of milk and milk products (Crielly, Logan, & Anderton, 1994). The use of ultrahigh-temperature (UHT) processing should inactivate the spores of *Bacillus* species and result in fluid milk products with a long shelf-life without refrigeration. However, defects in UHT milk product stability have been reported (Hammer, Lembke, Suhren, & Heeschen, 1995; Klijn et al., 1997). This nonstability appeared to be caused by the presence of highly heat-resistant bacterial endospores (HRS). HRS were first detected in UHT milk

from southern Europe in 1985 and were later identified in other countries both in and outside Europe (Hammer et al., 1995). They were found to belong to the species *Bacillus sporothermodurans*, first described by Pettersson, Lembke, Hammer, Stackebrand, and Fergus (1996). It thus became necessary to develop more efficient processes to inactivate HRS completely and ensure milk commercial sterility.

The combination of heat with high hydrostatic pressure (HP) has been proposed as an interesting approach for the inactivation of bacterial spores in food (Heinz & Buckow, 2010). HP has been shown to be efficient at destroying vegetative bacteria, and yeasts although bacterial spores appear to be more resistant to HP (Nakayama, Yano, Kobayashi, Ishikawa, & Sakai, 1996). San Martin, Barbosa-Canovas, and Swanson (2002) have shown that very high pressure levels (1000 MPa), at ambient temperature, are not effective in inactivating bacterial spores; however, this can be achieved by combining HP with temperatures higher than 40  C (Ju, Gao, Yao, & Qian, 2008; Kalchayanand, Dunne, Sikes, & Ray, 2004; Oh & Moon, 2003). Previous studies have concluded that

Abbreviations: DPA, Dipicolinic acid; HP, High pressure; HRS, Highly heat-resistant spores; RSM, Response surface methodology; UHT, Ultra-high temperature.

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a combination of pressure and moderate heat is always required to inactivate effectively spores of *Bacillus* species such as *Bacillus anthracis* (Cléry-Barraud, Gaubert, Masson, & Vidal, 2004), *Bacillus cereus* (Kannapha et al., 2008; Opstal, Bagamboula, Vanmuysen, Wuytack, & Michiels, 2004), *Bacillus subtilis* (Gao, Ju, & Jiang, 2006; Minh, Dantigny, Perrier-Cornet, & Gervais, 2010) and even *Clostridium* species (Kalchayanand et al., 2004; Mills, Earnshaw, & Patterson, 1998). For example, the inactivation rate of *B. anthracis* spores was shown to be higher (4.4-log cycle reduction) after a treatment at 280 MPa and 45 °C than after a treatment at 500 MPa and 20 °C (1-log cycle reduction) (Cléry-Barraud et al., 2004). Mills et al. (1998) showed that *Clostridium sporogenes* was resistant to HP, with treatments of 600 MPa for 30 min at 20 °C causing no significant inactivation, while the combination of mild heat (60 °C) with pressure (600 MPa for 30 min) resulted in a 3-log cycle reduction in spore count.

The mechanisms of spore-killing by heat, irradiation and chemicals have been well studied and described but those using HP are still poorly understood. It is now well known that HP can induce spore germination (Margosch, Ganzle, Ehrmann, & Vogel, 2004; Paidhungat et al., 2002; Wuytack, Boven, & Michiels, 1998; Wuytack, Soons, Poschet, & Michiels, 2000). Pressure (100–600 MPa) at 20–40 °C induces the opening of channels for the release of dipicolinic acid (DPA) from the spore core, which leads to spore germination (Paidhungat et al., 2002; Wuytack et al., 2000). Margosch et al. (2004) treated mashed carrots inoculated with spores of *B. subtilis*, *Bacillus amyloliquefaciens* and *Bacillus licheniformis* with HP and heat (200–800 MPa and 60–80 °C). Their results confirm that, at a pressure between 600 and 800 MPa and a temperature higher than 60 °C, DPA-free spores are inactivated by moderate heat independently of the pressure level.

The aim of this study was to evaluate the effectiveness of HP treatment, alone and in association with heat, on the inactivation of *B. sporothermodurans* spores in distilled water and in skim milk. For this purpose, response surface methodology (RSM) was used to describe spore inactivation in terms of pressure, temperature, and pressure-holding time.

## 2. Materials and methods

### 2.1. Bacterial strain

*B. sporothermodurans* strain LTIS27 was isolated from some contaminated samples of UHT-milk produced in Tunisia using the method previously described (Klijn et al., 1997). This strain was identified by PCR method using primers BSPO-F2 (5'-ACGGCTCAACCGTGAG-3') and BSPO-R2 (5'-GTAACCTCGCGGTCTA-3') (Biogene), as described by Scheldeman, Herman, Goris, De Voe, and Hendrickx (2002). The identification was confirmed by sequencing the 16S rRNA gene after PCR amplification as described by Winker and Woese (1991). *B. sporothermodurans* LTIS27 presents a high degree of similitude (99.8%) with the reference strain M215T type II regarding 16S rRNA gene sequences (Aouadhi et al., 2012), a specific strain of the UHT milk and named HRS clone (Montanari et al., 2004).

*B. sporothermodurans* LTIS27 was grown on brain–heart infusion agar supplemented with 1 mg/l vitamin B<sub>12</sub> (Sigma–Aldrich) (BHI-B<sub>12</sub>) at 37 °C. Twenty-four hours before use, cultures were held in BHI-B<sub>12</sub> broth on a rotary shaker at 37 °C to obtain a working culture.

### 2.2. Spore preparation

Spores were prepared according to the modified method of Ireland and Hanna (2002). A working culture (18 h, 37 °C under

**Table 1**

Codes and levels of factors chosen for the trials.

	Symbols		Level <sup>a</sup>				
	Coded	Uncoded	−1.68	−1.0	0.0	1.0	1.68
Pressure (MPa)	x <sub>1</sub>	X <sub>1</sub>	231.8	300	400	500	568.2
Temperature (°C)	x <sub>2</sub>	X <sub>2</sub>	23.8	30	40	50	56.2
Pressure-holding time (min)	x <sub>3</sub>	X <sub>3</sub>	3.2	10	20	30	36.8

$$^a x_1 = (X_1 - 400)/100; x_2 = (X_2 - 40)/10; x_3 = (X_3 - 20)/10.$$

agitation) was diluted 1:10 into sporulation medium (25 g l<sup>−1</sup> nutrient broth, 1 mg l<sup>−1</sup> vitamin B<sub>12</sub>, 8 mg l<sup>−1</sup> MnSO<sub>4</sub>·H<sub>2</sub>O, and 1 g l<sup>−1</sup> CaCl<sub>2</sub>·H<sub>2</sub>O) then incubated for 7 days at 37 °C. This medium was then centrifuged at 6000 g for 10 min. The resulting pellet, containing spores and non-sporulated vegetative cells, was washed three times with sterile distilled water, resuspended in 5 ml of sterile distilled water and treated at 100 °C for 30 min to kill the vegetative cells. The endospore suspension was centrifuged and washed 4 times as described above. The cleaned endospore preparation was stored at a final concentration of approximately 10<sup>7</sup>–10<sup>8</sup> spores ml<sup>−1</sup> in distilled water at 4 °C.

### 2.3. High hydrostatic-pressure treatment

High-pressure processing was carried out in a 3-l vessel (ACB Pressure Systems, Nantes, France) equipped with temperature and pressure regulatory devices, capable of achieving a maximum pressure of 600 MPa and a temperature of up to 60 °C. The samples (2 ml) were diluted 1:2 in sterile distilled water or in skim milk (5 × 10<sup>7</sup> spores ml<sup>−1</sup>), enclosed in polyethylene pouches, and subjected to the combined pressure and heat treatment protocols given in Table 2. The compression rate was 34 Pa/s and the pressure release was almost instantaneous. The temperature of the water in the vessel was maintained by circulating water within the double wall of the device from a water bath. The temperature increase due to the adiabatic heating effect was equivalent to approximately 1–3 °C/100 MPa depending on the initial temperature of the water in the vessel. After treatment, the pouches were immediately placed on ice and analyzed within 10 min.

### 2.4. Enumeration of surviving spores

Serial dilutions of treated samples were prepared in peptone water (0.01%) and the viable counts were determined by plating

**Table 2**

Central composite design arrangement for study the inactivation of *B. sporothermodurans* LTIS27 by pressure and moderate temperature.

Trial no.	Factors		
	X <sub>1</sub> (MPa)	X <sub>2</sub> (°C)	X <sub>3</sub> (min)
1	568.2	40.0	20.0
2	500.0	50.0	10.0
3	400.0	40.0	36.8
4	300.0	50.0	30.0
5	300.0	50.0	10.0
6	400.0	40.0	20.0
7	231.8	40.0	20.0
8	500.0	30.0	10.0
9	400.0	23.8	20.0
10	500.0	30.0	30.0
11	500.0	50.0	30.0
12	300.0	30.0	10.0
13	300.0	30.0	30.0
14	400.0	40.0	20.0
15	400.0	40.0	3.2
16	400.0	40.0	20.0
17	400.0	56.2	20.0

on BHI-B<sub>12</sub> agar. The plates were incubated at 37 °C for 24 h then the colonies were enumerated. Inactivation was expressed as a logarithmic viability reduction,  $\log(N_0/N)$ , with  $N$  and  $N_0$  the colony counts after treatment and in the untreated sample, respectively.

### 2.5. Experimental design

A rotatable central composite experimental design with three independent factors ( $\alpha = 1.68$ ) was used to study the inactivation of *B. sporothermodurans* spores in distilled water and in skim milk. The factors investigated were pressure, initial temperature in the vessel, and pressure-holding time. Each factor at five coded levels (indicated as -1.68, -1.0, 0.0, +1.0, +1.68 as shown in Table 1) were designed as experimental runs using the software STATGRAPHICS Centurion XV version 15.2.06. The real values of the independent variables ( $X$ ) were coded according to equation:  $x_i = (X_i - X_0)/\Delta X$ , Where  $X$  is the real value of the independent variable;  $x_i$  is the coded value of the independent variable given by the experimental matrix;  $X_0$  is the real value of the central point, and  $\Delta X$  is the step change.  $X$  and  $x$  values are shown in Table 1. The experimental response was the number of log cycle reduction of *B. sporothermodurans* spores ( $\log(N_0/N)$ ), which was estimated, taking into account the influence of the experimental factors. Three experiments were performed at the center of the experimental domain in order to estimate the residual variance value. An analysis of variance and an estimation of response surface by multiple linear regressions were performed using the software Statgraphics Centurion XV version 15.2.06.

## 3. Results

### 3.1. Predictive response model

Inactivation of *B. sporothermodurans* LTIS27 spores by the combined effects of pressure, temperature, and pressure-holding time in distilled water and in skim milk was studied, using RSM. The response ( $Y$ ) was measured in terms of log-cycle reduction ( $\log N_0/N$ ) ( $Y_1$ : in distilled water and  $Y_2$ : in skim milk). RSM is an empirical modeling technique used to estimate the relationship between the response variable and the test variable under consideration. This methodology has been used by some authors to evaluate the effect of various factors such as pressure-temperature on the inactivation of *B. cereus* (Ju et al., 2008) and *B. subtilis* (Gao & Jiang, 2005) spores. The response was represented by the following second-order polynomial equation, containing 10 estimated coefficients, where  $x_1$  is pressure,  $x_2$  is temperature, and  $x_3$  is pressure-holding time.

$$Y_1 = 15.69 - 0.052x_1 - 0.296x_2 - 0.159x_3 + 0.00004x_1^2 + 0.0002x_1x_3 + 0.0004x_1x_2 + 0.002x_2^2 - 0.0003x_2x_3 + 0.002x_3^2 \quad (1)$$

$$Y_2 = 9.82 - 0.033x_1 - 0.14x_2 - 0.149x_3 + 0.00003x_1^2 + 0.0002x_1x_3 + 0.0002x_1x_2 + 0.0016x_2^2 - 0.0001x_2x_3 + 0.002x_3^2 \quad (2)$$

The analysis of variance (ANOVA) of the quadratic model was performed with Statgraphics. The  $R^2$  values were 0.970 and 0.977 for Eqs. (1) and (2), respectively, indicating that the derived model fitted the experimental data. The statistical significance of each

coefficient value was determined using the Fisher  $F$ -test. The model  $F$ -value is the ratio of the mean square regression to the mean square residual. For the two Eqs. (1) and (2), the coefficient values were calculated and tested for their significance. The  $P$  values were used as a tool to check the significance of each coefficient which, in turn, may indicate the pattern of interactions between variables. The linear coefficients ( $x_1$ ,  $x_2$ , and  $x_3$ ), the quadratic term coefficients ( $x_1^2$ ,  $x_2^2$  and  $x_3^2$ ) and the cross coefficients ( $x_1x_2$  and  $x_1x_3$ ) were significant with small  $P$  values ( $P < 0.05$ ). The other term coefficient ( $x_2x_3$ ) was not significant.

### 3.2. Validation of the model

In order to validate the adequacy of polynomial equations (Eqs. (1) and (2)), ten experimental checks were carried out under different HP combinations of process parameters (Table 3). The experimental values were found to be significantly in agreement with the predicted values with a correlation coefficient ( $R^2$ ) of 0.99 and a statistical significance level of  $P < 0.0001$ . Therefore, the model was proven to be quite reliable for predicting the inactivation of *B. sporothermodurans* spores in distilled water and in skim milk for treatment parameters in the range of 231.8–568.2 MPa, 23.8–56.2 °C and 3.2–36.8 min.

### 3.3. Response surfaces

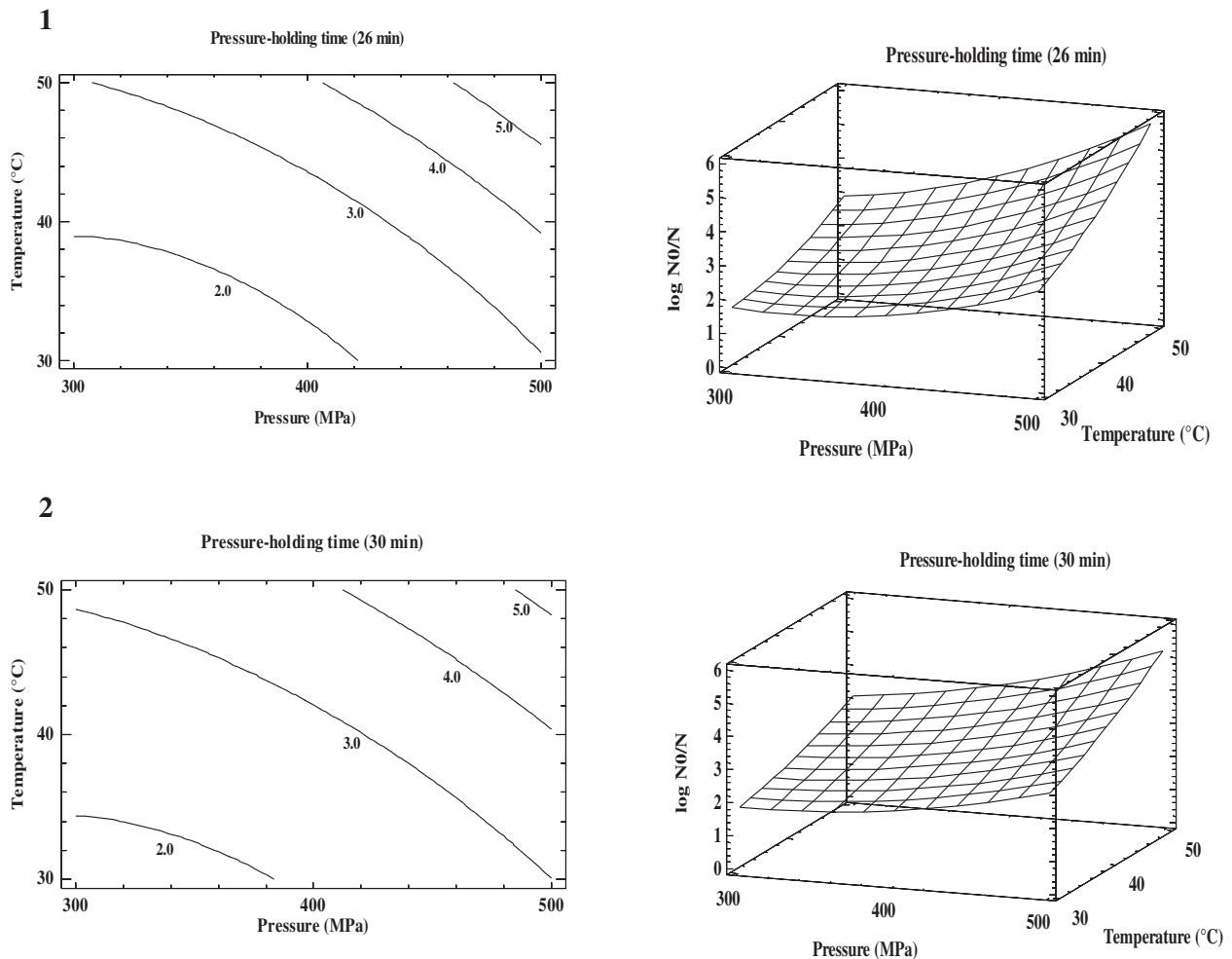
Spores of *B. sporothermodurans* are known to be extremely resistant to heat treatment. The maximal load of this bacteria reported in stored milk is  $10^5$  cfu ml<sup>-1</sup> of milk after 5 days of storage at 30 °C (Hammer et al., 1995; Klijn et al., 1997). Thus, the process parameters for a 5-log cycle reduction of *B. sporothermodurans* spores in distilled water and in skim milk were calculated using the model. The graphical representations were obtained by solving the regression Eqs. (1) and (2) using Statgraphics. In Figs. 1–3, response surfaces and contour plots show the effect of pressure, temperature and pressure-holding time on the inactivation of spores. In each figure, one factor is maintained constant at its optimal value determined by Statgraphics. Fig. 1 shows the effect of pressure and temperature while keeping the pressure-holding time at its optimal value (26 min in distilled water and 30 min in skim milk). The reduction of *B. sporothermodurans* spores increases with increasing pressure and temperature, reaching a 5-log cycle reduction at pressures of 460–500 MPa and temperatures of 45–50 °C.

The interaction of pressure and pressure-holding time at a constant temperature (48 °C in distilled water and 49 °C in skim milk) is shown in Fig. 2. In the pressure range of 480–500 MPa and

**Table 3**

Experimental and predicted results for ten check-points of inactivation of *B. sporothermodurans* LTIS27 spores in distilled water and in milk.

Trial no.	Factors			$Y = \log N_0/N$			
	$X_1$ (MPa)	$X_2$ (°C)	$X_3$ (min)	In distilled water		In skim milk	
				Observed	Predicted	Observed	Predicted
1	320	40	10	1.9	1.8	1.6	1.5
2	350	50	15	2.8	2.7	2.4	2.4
3	370	30	10	1.1	1.2	0.9	0.8
4	390	45	20	2.6	2.5	2.3	2.2
5	420	40	30	2.9	2.9	2.4	2.5
6	450	35	23	2.3	2.2	1.9	1.8
7	490	45	16	3.2	3.3	2.8	2.9
8	520	40	18	3.3	3.2	2.8	2.7
9	523	47	30	5.2	5.2	4.6	4.7
10	530	48	30	5.4	5.5	5.0	5.0



**Fig. 1.** Response surface and contour plots showing the effects of pressure and temperature on the inactivation of *B. sporothermodurans* LTIS27 spores. The data on the contour maps are  $\log N_0/N$  in distilled water (1) and milk (2).  $N_0$  represents the plate count of the untreated spore suspension ( $5 \times 10^7$  spore  $\text{ml}^{-1}$ ) and  $N$  represents the plate count after pressure treatment.

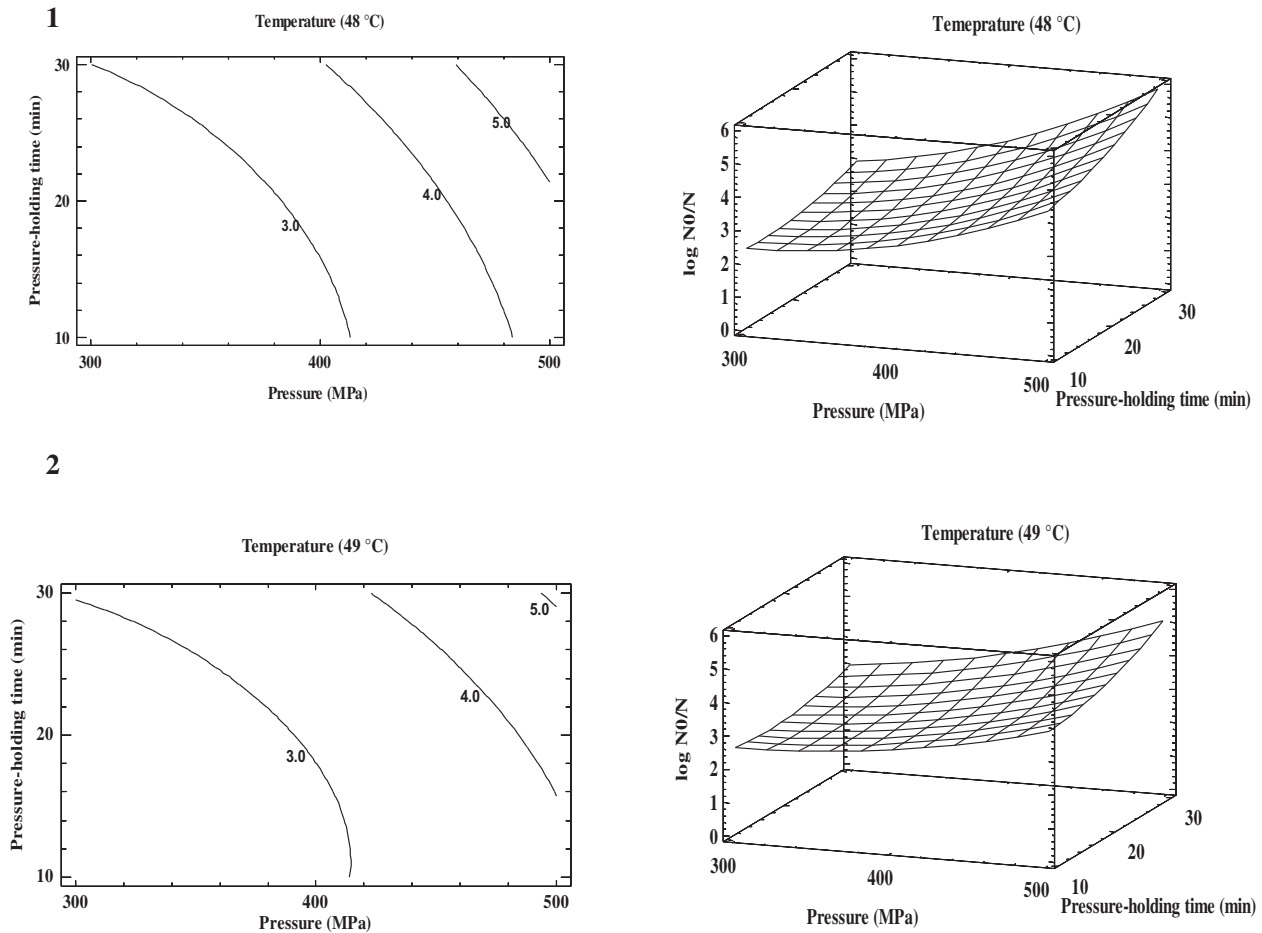
the pressure-holding time range of 25–30 min, a reduction of 5-log cycle of *B. sporothermodurans* spores was calculated. The reduction of *B. sporothermodurans* spores as a function of temperature and pressure-holding time at a fixed pressure of 477 MPa in distilled water and 495 MPa in skim milk is given by the contour plots in Fig. 3. The reduction of 5-log cycle of *B. sporothermodurans* spores was obtained in the temperature range of 47–50 °C and in the pressure-holding time range of 24–30 min.

Optimum values of each parameter given by the model are as follows: 477 MPa/48 °C for 26 min in distilled water and 495 MPa/49 °C for 30 min in skim milk.

#### 4. Discussion

The inactivation of *B. sporothermodurans* spores by combined HP and heat treatment was more efficient in distilled water than in skim milk. The process conditions leading to a 5-log cycle reduction of *B. sporothermodurans* spores were calculated using the model and less drastic conditions were obtained in water (477 MPa/48 °C for 26 min) than in milk (495 MPa/49 °C for 30 min). Some constituents of milk may protect spores against HP. The protective effect of food constituents on vegetative bacterial cells is well known and documented. Several authors have reported that bacteria are more resistant in a complex matrix such as milk or

meat (Black, Huppertz, Fitzgerald, & Kelly, 2007; Chen & Hoover, 2003; Kannapha et al., 2008). Chen and Hoover (2003) demonstrated a strong baroprotective effect of whole UHT milk on *Yersinia enterocolitica*, with inactivation levels 3.5–4.5-log cycle lower than in phosphate buffer when treated at 350–450 MPa/22 °C for 10 min Black et al. (2007) studied the baroprotective effect of milk constituents on *Listeria innocua* treated by HP and stated that the divalent cations, calcium and magnesium, may protect cell membranes against HP. The effect of the food matrix on spore inactivation is more contradictory in the literature. While Kannapha et al. (2008) showed that crab meat had a protective effect against inactivation of *B. cereus* spores in comparison with spores suspended in distilled water treated at 550 MPa/40 °C for 15 min, Opstal et al. (2004) reported that the inactivation of *B. cereus* spores was 1-log cycle reduction higher in milk than in phosphate buffer after a treatment at 600 MPa, 60 °C for 10 min. The inactivation level of spores does not always follow the trend of germination since this was shown to be enhanced in crab meat (Kannapha et al., 2008) and in milk (Aouadhi et al., 2012; Opstal et al., 2004) compared to distilled water or buffer. The increase in germination in milk or in crab meat was attributed to the presence of some germinant, such as L-alanine, required in the germination process in addition to pressure. Thus, it seems that food nutrients have two opposite effects on spores under pressure; some may



**Fig. 2.** Response surface and contour plots showing the effects of pressure and pressure-holding time on the inactivation of *B. sporothermodurans* LTIS27 spores. The data on the contour maps are  $\log N_0/N$  in distilled water (1) and milk (2).  $N_0$  represents the plate count of the untreated spore suspension ( $5 \times 10^7$  spore  $\text{ml}^{-1}$ ) and  $N$  represents the plate count after pressure treatment.

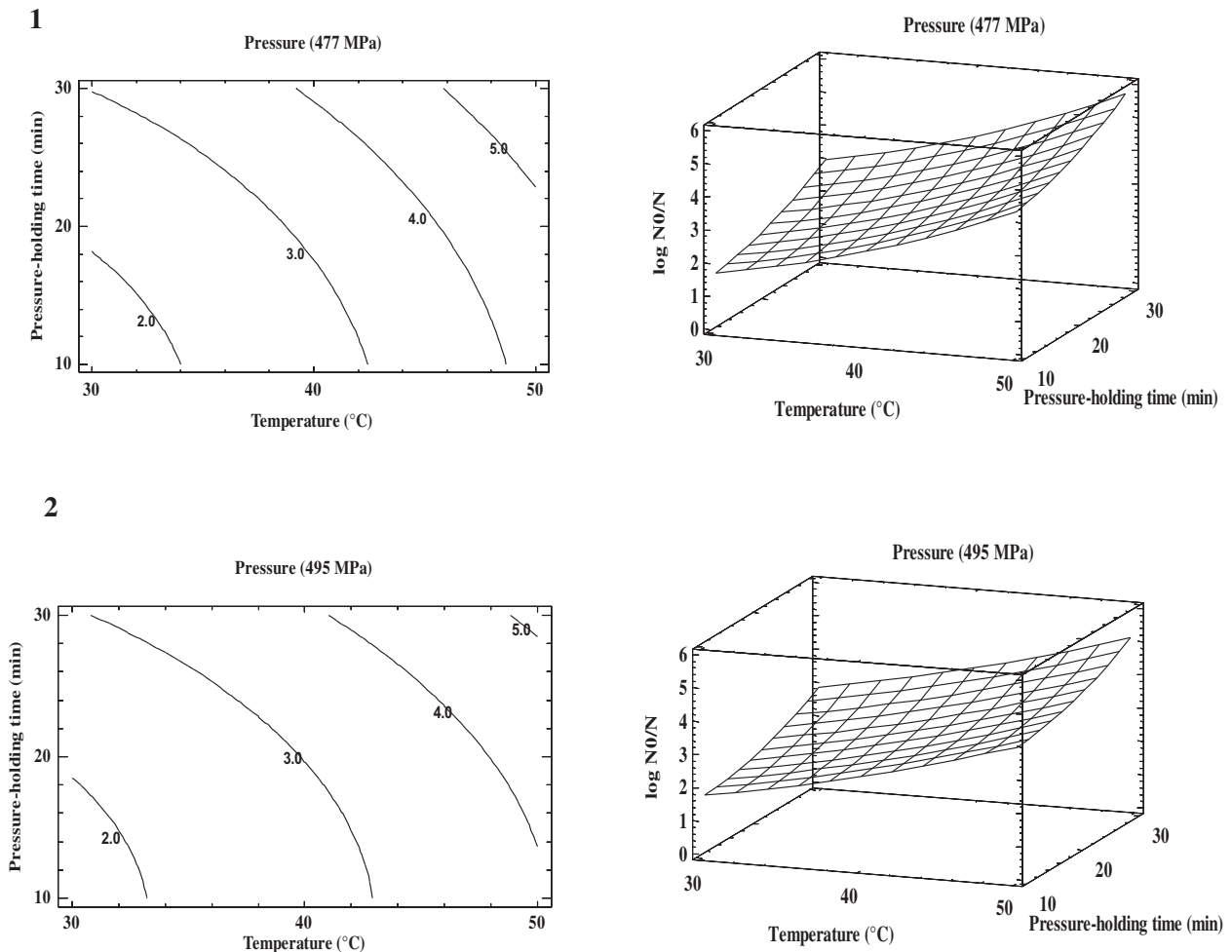
accelerate germination while others may protect germinated spores against inactivation. In the present work, the protective effect of milk constituents against thermal inactivation of DPA-free spores could predominate.

Whatever the suspension medium, spore inactivation increased as the three factors of pressure, temperature and pressure-holding time increased. The model showed significant factor interaction meaning that significant synergistic effects occur with pressure–temperature and pressure–pressure-holding time in distilled water and in milk. At temperatures higher than 40 °C, the treatment pressure and pressure-holding time may be decreased to obtain the same inactivation level. In the same way, for pressure levels higher than 400 MPa, the treatment temperature and the processing time may be decreased to provide the same inactivation level. Nevertheless, significant inactivation requires minimal values of pressure and temperature while the pressure-holding time has only a significant effect for pressure treatments above 400 MPa and above 40 °C. Thus, the treatment conditions must be severe enough both to induce the germination process and to inactivate the germinated spores.

Several authors have stated that a considerable (more than 5-log cycle) and reproducible reduction of spores by pressure of around 600 MPa is only possible in combination with temperatures between 75 and 80 °C (Cléry-Barraud et al., 2004; Gao et al., 2006; Margosch et al., 2004). For example, the optimum process parameters for a 6-log cycle reduction of *Geobacillus stearothermophilus*

were calculated using RSM to be 625 MPa, 86 °C and 14 min (Gao et al., 2006). Moerman, Mertens, Demey, and Huyghebaert (2001) demonstrated a 5–6-log cycle reduction of *Geobacillus stearothermophilus* spores after processing at 400 MPa/80 °C for 60 min. According to Gao et al. (2006), a 6-log cycle reduction of *B. subtilis* spores in milk was obtained with a treatment at 576 MPa, 87 °C for 13 min. The present study shows that, for *B. sporothermodurans* LTIS27 spores, a 5-log cycle inactivation can be obtained with less drastic treatment conditions of 477 MPa/48 °C for 26 min and 495 MPa/49 °C for 30 min, in distilled water and in milk, respectively.

*B. sporothermodurans* spores are known to be highly heat-resistant with  $D_{140}$  values ranging from 3.4 to 7.9 s in skim milk (Huemer, Klijn, Vogelsang, & Langeveld, 1998). Using the data obtained by Moerman et al. (2001), we compared the inactivation of *B. sporothermodurans* in milk with the inactivation of other highly heat-resistant spores, the spores of *G. stearothermophilus* suspended in fatty meat. After the same treatment at 400 MPa, 50 °C for 30 min, 1.52 and 3.68-log cycle were obtained with the model developed by Moerman et al. (2001) on *G. stearothermophilus* inactivation and the model of the present study on *B. sporothermodurans*, respectively. Although the compared results concern only one specific strain of each bacteria, this could mean that the heat resistance of *B. sporothermodurans* spores has no relation to its pressure resistance. In the literature, two opinions coexist about the relationship between the pressure and heat



**Fig. 3.** Response surface and contour plots showing the effects of temperature and pressure-holding time on the inactivation of *B. sporothermodurans* LTIS27 spores. The data on the contour maps are  $\log N_0/N$  in distilled water (1) and milk (2).  $N_0$  represents the plate count of the untreated spore suspension ( $5 \times 10^7$  spore  $\text{ml}^{-1}$ ) and  $N$  represents the plate count after pressure treatment.

resistance of spores. Rocken and Spicher (1993) demonstrated that the highest resistance to pressure was observed in highly heat-resistant strains of *B. subtilis* and *B. amyloliquefaciens* previously isolated from ropy bread. This finding may indicate a correlation between heat resistance and pressure resistance. However, spores of *B. amyloliquefaciens* are considerably more pressure-resistant than spores of *G. stearothermophilus*, which exhibit a higher resistance to wet heat (Ananta, Heinz, Schluter, & Knorr, 2001). Furthermore, Nakayama et al. (1996), by comparing the pressure and heat resistance of spores from six *Bacillus* strains, indicated that, for some strains, the pressure resistance did not correlate well with the heat resistance values obtained. Therefore, no pressure treatment condition could be converted into a heat sterilization condition. The sporulation medium, pressure-transmitting fluid, initial treatment temperature, and the high pressure equipment may all have a strong influence on the resistance of *Bacillus* spores to combined HP-heat treatments (Nakayama et al., 1996).

Clearly, the HP-resistance of *Bacillus* species is highly variable. Margosch et al. (2004), while studying the levels of resistance of 14 food isolates and 5 laboratory strains of *Bacillus* species to combined pressure and temperature treatments, found a large variation in the pressure resistance of spores. The reduction by treatment at 800 MPa and 70 °C for 4 min ranged from more than 6 log units for *B. subtilis* and *B. licheniformis* to no reduction for *B. amyloliquefaciens*. For vegetative cells, it has been reported that

different microbial species, and even strains within species, can exhibit different HP sensitivity (Patterson, Quinn, Simpson, & Gilmour, 1995). This may also be true for bacterial spores. Furthermore, HP-induced spore inactivation depends on a range of factors, related to the microorganism itself, the conditions of treatment (levels of pressure, treatment time, and temperature) and to the medium in which the microorganisms are suspended. Recently, Minh, Durand, Loison, Perrier-Cornet, and Gervais (2011) also showed that the sporulation conditions may affect the HP-resistance of spores. Such variability makes the modeling of the HP-inactivation of spores a very difficult challenge.

## 5. Conclusion

RSM involving experimental design and regression analysis was used to reveal the influence of pressure, temperature, and pressure-holding time on the inactivation of *B. sporothermodurans* spores in distilled water and in skim milk. Our results suggest that the association of heat with pressure is very efficient for the inactivation of *B. sporothermodurans*, as has been shown for other spore-forming species, more so in distilled water than in milk. The optimum process parameters obtained for a 5-log cycle reduction of *B. sporothermodurans* spores were 477 MPa/48 °C for 26 min in distilled water and 495 MPa/49 °C for 30 min in milk. This study shows the potential of using HP in combination with moderate

temperature to inactivate *B. sporothermodurans* spores in milk. It is particularly interesting because the initial contamination density ( $N_0$ ) of HRS spores in raw milk is generally lower than  $10^5$  cfu ml<sup>-1</sup>. However, for scale-up process validation, a multiple-strain study should be performed. Finally, this work opens up new perspectives for the study of *B. sporothermodurans* spore behavior under HP.

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### References

- Ananta, E., Heinz, V., Schluter, O., & Knorr, D. (2001). Kinetic studies on high-pressure inactivation of *Bacillus stearothermophilus* spores suspended in food matrices. *Innovative Food Science and Emerging Technologies*, 2, 261–272.
- Aouadhi, C., Simonin, H., Prevost, H., De Lamballerie, M., Maaroufi, A., & Mejri, S. (2012). Optimization of pressure-induced germination of *Bacillus sporothermodurans* spores in water and milk. *Food Microbiology*, 30, 1–7.
- Black, E. P., Huppertz, T., Fitzgerald, G. F., & Kelly, A. L. (2007). Baroprotection of vegetative bacteria by milk constituents: a study of *Listeria innocua*. *International Dairy Journal*, 17, 104–110.
- Chen, H., & Hoover, D. G. (2003). Pressure inactivation kinetics of *Yersinia enterocolitica* ATCC 35669. *International Journal of Food Microbiology*, 87, 161–171.
- Cléry-Barraud, C., Gaubert, A., Masson, P., & Vidal, D. (2004). Combined effects of high hydrostatic pressure and temperature for inactivation of *Bacillus anthracis* spores. *Applied and Environmental Microbiology*, 70, 635–637.
- Crielly, E. M., Logan, N. A., & Anderton, A. (1994). Studies on the *Bacillus* flora of milk and milk products. *Journal of Applied Bacteriology*, 77, 256–263.
- Gao, Y., & Jiang, H. (2005). Optimization of process conditions to inactivate *Bacillus subtilis* spores by high hydrostatic pressure and mild heat using response surface methodology. *Biochemical Engineering Journal*, 24, 43–48.
- Gao, Y., Ju, X., & Jiang, H. (2006). Analysis of reduction of *Geobacillus stearothermophilus* spores treated with high hydrostatic pressure and mild heat in milk buffer. *Journal of Biotechnology*, 125, 351–360.
- Hammer, P., Lembke, F., Suhren, G., & Heesch, W. (1995). Characterization of a heat resistant mesophilic *Bacillus* species affecting quality of UHT-milk – a preliminary report. *Kiel Milchwirtschafts Forschungsber*, 47, 303–311.
- Heinz, V., & Buckow, R. (2010). Food preservation by high pressure. *Journal of Consumer Protection and Food Safety*, 5, 73–81.
- Huemer, I. A., Klijn, N., Vogelsang, H. W. J., & Langeveld, L. P. M. (1998). Thermal death kinetics of spores of *Bacillus sporothermodurans* isolated from UHT-milk. *International Dairy Journal*, 8, 851–855.
- Ireland, J. A., & Hanna, P. C. (2002). Amino acid- and purine ribonucleoside- induced germination of *Bacillus anthracis* Å Sterne endospores: *gerS* mediates responses to aromatic ring structures. *Journal of Bacteriology*, 84, 1296–1303.
- Ju, X. R., Gao, Y. L., Yao, M. L., & Qian, Y. (2008). Response of *Bacillus cereus* spores to high hydrostatic pressure and moderate heat. *Food Science and Technology*, 41, 2104–2112.
- Kalchayanand, N., Dunne, C. P., Sikes, A., & Ray, B. (2004). Germination induction and inactivation of *Clostridium* spores at medium-range hydrostatic pressure treatment. *Innovative Food Science and Emerging Technologies*, 5, 277–283.
- Kannapha, S., George, J. F., Dianne, W. B., Ankenman, L. G., Joseph, E., & Robert, W. (2008). Pressure-induced germination and inactivation of *Bacillus cereus* spores and their survival in fresh blue crab meat (*Callinectes sapidus*) during storage. *Journal of Aquatic Food Product Technology*, 17, 322–337.
- Klijn, N., Herman, L., Langeveld, L., Vaerewijck, M., Wagendorp, A., & Huemer, I. (1997). Genotypical and phenotypical characterization of *Bacillus sporothermodurans* strains, surviving UHT sterilization. *International Dairy Journal*, 7, 421–428.
- Margosch, D., Ganzle, M. G., Ehrmann, M. A., & Vogel, R. F. (2004). Pressure inactivation of *Bacillus* endospores. *Applied and Environmental Microbiology*, 70, 7321–7328.
- Mills, G., Earnshaw, R., & Patterson, M. F. (1998). Effects of high hydrostatic pressure on *Clostridium sporogenes* spores. *Letters in Applied Microbiology*, 26, 227–230.
- Minh, H. N. T., Dantigny, P., Perrier-Cornet, J. M., & Gervais, P. (2010). Germination and inactivation of *Bacillus subtilis* spores induced by moderate hydrostatic pressure. *Biotechnology and Bioengineering*, 107, 876–883.
- Minh, H. N. T., Durand, A., Loison, P., Perrier-Cornet, J. M., & Gervais, P. (2011). Effect of sporulation conditions on the resistance of *Bacillus subtilis* spores to heat and high pressure. *Applied Microbiology and Biotechnology*, 90, 1409–1417.
- Moerman, F., Mertens, B., Demey, L., & Huyghebaert, A. (2001). Reduction of *Bacillus subtilis*, *Bacillus stearothermophilus* and *Streptococcus faecalis* in meat batters by temperature-high hydrostatic pressure pasteurization. *Meat Science*, 59, 115–125.
- Montanari, G., Borsari, A., Chiavari, C., Ferri, G., Zambonelli, C., & Grazia, L. (2004). Morphological and phenotypical characterization of *Bacillus sporothermodurans*. *Journal of Applied Microbiology*, 97, 802–809.
- Nakayama, A., Yano, Y., Kobayashi, S., Ishikawa, M., & Sakai, K. (1996). Comparison of pressure resistances of spores of six *Bacillus* strains with their heat resistances. *Applied and Environmental Microbiology*, 62, 3897–3900.
- Oh, S., & Moon, M. (2003). Inactivation of *Bacillus cereus* spores by high hydrostatic pressure at different temperatures. *Journal of Food Protection*, 66, 599–603.
- Opstal, I. V., Bagamboula, C. F., Vanmuysen, S. C., Wuytack, E. Y., & Michiels, C. W. (2004). Inactivation of *Bacillus cereus* spores in milk by mild pressure and heat treatments. *International Journal of Food Microbiology*, 92, 227–234.
- Paidhungat, M., Setlow, B., Daniels, W. B., Hoover, D., Papafragkou, E., & Setlow, P. (2002). Mechanisms of induction of germination of *Bacillus subtilis* spores by high pressure. *Applied and Environmental Microbiology*, 68, 3172–3175.
- Patterson, M. F., Quinn, M., Simpson, R., & Gilmour, A. (1995). Sensitivity of vegetative pathogens to high hydrostatic pressure treatment in phosphate-buffered saline and foods. *Journal of Food Protection*, 58, 524–529.
- Pettersson, B., Lembke, F., Hammer, P., Stackebrand, E., & Fergus, G. (1996). *Bacillus sporothermodurans* a new species producing highly heat resistant endospores. *International Journal of Systematic Bacteriology*, 46, 759–764.
- Rocken, W., & Spicher, G. (1993). Fadenziehende Bakterien-Vorkommen, Bedeutung Gegenmaßnahmen. *Getreide Mehl Brot*, 47, 30–35.
- San Martin, M. F., Barbosa-Canovas, G. V., & Swanson, B. G. (2002). Food processing by high hydrostatic pressure. *Critical Reviews in Food Science and Nutrition*, 42, 627–645.
- Scheldeman, P., Herman, L., Goris, J., De Vo, P., & Hendrickx, M. (2002). Polymerase chain reaction identification of *Bacillus sporothermodurans* from dairy source. *Journal of Applied Microbiology*, 92, 983–991.
- Winker, S., & Woese, C. R. (1991). A definition of the domains *Archaea*, *Bacteria* and *Eucarya* in terms of small subunit ribosomal rRNA characteristics. *Systematic Applied Microbiology*, 13, 161–165.
- Wuytack, E. Y., Boven, S., & Michiels, C. W. (1998). Comparative study of pressure-induced germination of *Bacillus subtilis* spores at low and high pressures. *Applied and Environmental Microbiology*, 64, 3220–3224.
- Wuytack, E. Y., Soons, J., Poschet, F., & Michiels, C. W. (2000). Comparative study of pressure- and nutrient-induced germination of *Bacillus subtilis* spores. *Applied and Environmental Microbiology*, 66, 257–261.